Emerging Concepts in Alzheimer's Disease

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Abstract

Alzheimer's disease/senile dementia of the Alzheimer type (AD/SDAT) is the most common neuropathologic substrate of dementia. It is characterized by synapse loss (predominantly within neocortex) as well as deposition of certain distinctive lesions (the result of protein misfolding) throughout the brain. The latter include senile plaques, composed mainly of an amyloid (Aβ) core and a neuritic component; neurofibrillary tangles, composed predominantly of hyperphosphorylated tau; and cerebral amyloid angiopathy, a microangiopathy affecting both cerebral cortical capillaries and arterioles and resulting from AB deposition within their walls or (in the case of capillaries) immediately adjacent brain parenchyma. In this article, I discuss the hypothesized role these lesions play in causing cerebral dysfunction, as well as CSF and neuroimaging biomarkers (for dementia) that are especially relevant as immunotherapeutic approaches are being developed to remove Aß from the brain parenchyma. In addition, I address the role of neuropathology in characterizing the sequelae of new AD/SDAT therapies and helping to validate CSF and neuroimaging biomarkers of disease. Comorbidity of AD/SDAT and various types of cerebrovascular disease is a major theme in dementia research, especially as cognitive impairment develops in the oldest old, who are especially vulnerable to ischemic and hemorrhagic brain lesions.

INTRODUCTION

AD: Alzheimer's disease

SDAT: senile dementia of Alzheimer type

Aβ: amyloid β

MRI: magnetic resonance imaging

PET: positron emission tomography

Dementia is a clinically defined entity or syndrome; though it has many causes, the most common etiology of dementia is Alzheimer's disease (1–5). Alzheimer's disease/senile dementia of the Alzheimer type (AD/SDAT) is the most common neurodegenerative disease worldwide; it afflicts over 5 million Americans, and given that aging is the major risk factor for developing it, absent a cure or definitive prevention strategy its incidence will continue to increase in the coming years as medical advances lead to increasing life span (6). One estimate based on US Census Bureau data is that, between 2000 and 2020, the number of individuals living to 100 years or more will increase by over 200% and the number of individuals surviving to 90–95 years will double (7). From an epidemiologic perspective, the Baltimore Longitudinal Study of Aging (1985–1998) found that the incidence rates of AD increased with age from an estimated 0.08% per year in the 60–65-year age group to an annual estimated incidence of 6.48% in the \geq 85-year age group (8). This study furthermore estimated that the doubling time of incidence rates was approximately 4.4 years, with a similar time interval for conversion from mild cognitive impairment (MCI) to AD.

Autopsy of a subject with this clinical diagnosis is used to confirm the clinical assessment and to assess any comorbidities that may have contributed to cognitive impairment (2, 5, 9, 10). The neuropathologic diagnosis of AD in a given subject is made by evaluating the location, distribution, and abundance of characteristic brain lesions. Attempts to standardize the neuropathologic diagnosis of AD and comorbidities have evolved rapidly beginning in the 1980s and continuing to the present (see below). Neurodegenerative diseases are traditionally considered as disorders in which progressive loss of neurons and synapses occurs in distinct anatomical loci, resulting in different phenotypes. A defining biochemical theme in the study of most neurodegenerative disorders is that of protein misfolding, the molecular nature of which is explored in depth (and from various perspectives) in a recent book (11). In this review I discuss the practical aspects of gross, microscopic, and immunohistochemical features of AD/SDAT; how the neuropathologist optimally approaches evaluation of an autopsy or biopsy brain specimen from a clinically affected individual; and how selected biochemical, molecular, and genetic findings—of which there has been an explosion in recent years—inform and often refine the clinicopathologic evaluation of brains originating from cognitively impaired individuals or from those lacking neurologic impairment.

Sensitive neuroimaging techniques are emerging and being refined that are capable of both quantifying AD-associated cerebral atrophy and detecting amyloid β (A β) peptide, phosphorylated tau (phospho-tau), or other β -pleated sheet proteins in the brain while patients are alive, even when completely asymptomatic or at early stages of neurodegeneration (12–15; see also the discussion of biomarkers below). Yet detailed autopsy-or (less commonly) biopsy neuropathologic examination of the brain-continues to be considered the gold standard for the diagnosis of AD and non-AD dementias (1, 2, 5, 16). Careful clinicopathologic correlation, that is, attempting to explain complex neurologic symptoms in an often end-stage central nervous system (CNS) by autopsy examination of the brain, was a central pillar of dementia research through the 1970s. At the end of the 1970s, structural imaging emerged and began to provide valuable information about the CNS in vivo, though CT scanning was most valuable for assessing mass lesions in the brain but was less effective at detecting cortical atrophy. Magnetic resonance imaging (MRI) and positron emission tomography (PET), the latter performed with highly specialized ligands, have provided valuable information in the study of neurodegenerative disorders (see below). It bears reemphasis that the starting material for important biochemical and molecular studies that have linked abnormally folded proteins to neurodegeneration was rapidly harvested (autopsy) brain tissue, the neuropathologic features of which were subsequently often correlated with the relevant neurochemical data (1, 4, 17, 18).



Coronal slices of (fixed) brain from two different patients, one without dementia (panel *a*) and one with (panel *b*); slices are at comparable coronal levels (near the head of the caudate nucleus). Arrows indicate a relatively normal lateral ventricle in the control case (panel *a*) versus a markedly enlarged lateral ventricle in the Alzheimer's disease subject (panel *b*). Cortical thinning is less prominent.

The main neuropathologic feature of the AD brain on gross inspection is cortical atrophy, which is usually diffuse and fairly symmetrical throughout the cerebral hemispheres rather than being accentuated in certain lobes or on one side of the brain [as in the case of some frontotemporal lobar degenerations (FTLDs)] (1, 2, 5, 19-21). Fresh brain weight is usually below the normal range for an adult (1,200–1,400 g), though not necessarily so—it may be entirely normal and even, rarely, above the upper range of normal (2, 5, 22). When the fixed brain is sliced, the cortical atrophy (manifest as thinning of the cortical ribbon) is usually accompanied by enlargement of the ventricular system, or hydrocephalus ex vacuo, and sometimes by shrinkage, atrophy, and/or pallor of the subcortical white matter (Figure 1). Precise etiology of the white matter change is not known—it may in part represent downstream (anterograde, Wallerian) degeneration secondary to cortical atrophy with neuron loss or may be the manifestation of the leukoencephalopathy that is an integral component of AD/SDAT (23). If the brain of a demented patient shows hydrocephalus out of proportion to the degree of cerebral cortical atrophy, the possible diagnosis of normal pressure hydrocephalus must be considered, though microscopic lesions of AD should still be sought in such a brain. Most experienced neuropathologists (myself included) are impressed by the variability in brain weights, cerebral cortical atrophy, and hydrocephalus ex vacuo among individuals who eventually have the diagnosis of AD confirmed by light microscopy (22).

ALZHEIMER'S DISEASE: MICROSCOPIC LESIONS

As Nelson et al. have emphasized (7, 24), AD/SDAT is defined by the presence of three elements one clinical and the other two neuropathologic:

- clinical dementia (cognitive impairment, with a memory component, that impacts daily living skills);
- 2. substantial numbers of neurofibrillary tangles (NFTs) within neocortex; and
- 3. significant numbers of senile (neuritic) plaques within the brain.

(The issue of whether AD can be diagnosed in the brain of someone who has not experienced significant well-documented neurocognitive abnormalities is addressed below.) The microscopic lesions that are known to accumulate in the CNS (mainly cerebral cortex, but often also deep central gray matter, cerebellum, and brainstem) of individuals with AD can, when prominent and numerous, be identified on routine sections of the brain; however, they are much more easily demonstrated

NFT: neurofibrillary tangle



Senile plaques (SPs). (*a*, *b*) Hematoxylin and eosin (H&E)-stained sections show a neuritic SP without a prominent amyloid core (panel *a*, *arrows*) and two SPs with prominent amyloid cores (panel *b*, *arrows*). (*c*) Section immunostained with phospho-tau shows an SP with a prominent tau-immunoreactive neuritic component (*arrows*). Note tau-immunoreactive neurofibrillary tangles (or pretangles) in the surrounding cortex.

by the use of special stains, especially silver impregnation techniques (modified Bielschowsky, Bodian, Campbell–Switzer, and Gallyas techniques) (1, 2, 5, 9, 25–27) and immunohistochemical methods incorporating primary antibodies to proteins deposited within afflicted brain. Though many of the older silver stains have an intrinsic elegance, they are sometimes capricious and result in annoying tissue section artifacts that tend to limit their usefulness.

SENILE PLAQUES

Neuritic senile plaques (SPs) appear, on routine hematoxylin & eosin (H&E)-stained sections, as an irregularity or coarsening of the neuropil (the neuritic component of the SP) centered on the core of the SP, an amorphous eosinophilic globule of amyloid (**Figure 2**). Masters et al. (28) showed that SP cores are composed of A β . The dynamic relationship between the amyloid core of a mature SP and its neuritic corona (both seen on silver stains) has been debated for years, but such mature neuritic SPs are thought to be more reflective of cortical brain injury than are the more diffuse SPs lacking a neuritic component (1, 2). Diffuse SPs (seen using silver stains or A β immunohistochemistry) are a common finding in the brains of elderly cognitively normal subjects (7, 24). Outstanding reviews on the hypothesized cellular and molecular pathogenesis of SPs are available (e.g., 29). The precise role of astrocytes and microglia in SP genesis [through the generation of A β from amyloid precursor protein (APP)] or A β clearance is controversial. Although neuritic SPs have a neuronal component, insofar as the neurites encircling the amyloid core represent processes emerging from (presumably) damaged nerve cell bodies, they are located within the neuropil. Although SPs are often found in elderly individuals without AD, their density among

SP: senile plaque **APP:** amyloid precursor protein

such subjects is far less than that in patients with AD (30); most neuropathologists have, however, encountered autopsy brains from cognitively intact elderly individuals that contain abundant neuritic SPs. Anecdotal reports have described all neuropathologic features of AD in cognitively normal elderly—indeed, even in rare individuals who had been examined neurologically shortly before death and judged to be cognitively intact (7, 31). Dense core SPs may be identified with Thioflavin S stains (32). One study has shown that SPs vary in size among both AD patients and normal elderly individuals; larger size distributions correlated with an earlier age of disease onset. SPs did not, however, appear to increase significantly in size over the clinical course of the disease (32). Thal et al. (33) have proposed five neuroanatomical phases in the deposition of A β within the CNS: deposition in the neocortex, allocortex, diencephalic-striatal-basal forebrain, brainstem, and cerebellum, with the final stage representing the most advanced A β pathology.

NEUROFIBRILLARY TANGLES

NFTs are dense intraneuronal cytoplasmic basophilic aggregates of filaments that include, on ultrastructural examination, characteristic paired helical filaments (PHFs; also described as bifilar helices) (1, 2, 5, 34). PHFs were first identified by Kidd (35) in brain biopsies; the filaments have a diameter of 10–15 nm and a periodicity of 160 nm. NFTs are almost invariably accompanied by neuropil threads in the adjacent brain parenchyma—these threads are interpreted as constituting the processes of NFT-bearing neurons (36, 37). Tangles tend to accommodate to the shape and morphology of a given neuron; when present in the pyramidal cell layer of the hippocampus, they appear as elongated parallel skeins of fibrillar intracytoplasmic basophilic material, but when seen in globose or oval neurons, they take on a more disorganized or globular configuration. A provocative study of more than 2,330 autopsy brains over an age range of 1–100 years (27) concluded that abnormal intraneuronal tau deposits may occur as early as the second or third decade of life (at least within the transentorhinal cortex) and that brainstem tau deposition (e.g., in the locus ceruleus) may even antedate transentorhinal changes.

NFTs (Figure 3) occur in the CNS of patients with many non-AD neurodegenerative conditions, including subacute sclerosing panencephalitis, postencephalitic parkinsonism, dementia pugilistica, and the parkinsonian–amyotrophic lateral sclerosis–dementia complex of Guam (38). NFT-like neuronal cytoplasmic lesions are commonly encountered within the dysmorphic and enlarged neuronal cell bodies of infants and children with epilepsy-causing cortical dysplasia or



Figure 3

Neurofibrillary tangles (NFTs). Sections were immunostained with primary antibody to phospho-tau and visualized at low (panel *a*) and high (panel *b*) magnification. Note that most neurons in the field show tau immunoreactivity. Arrows in panel *b* highlight immunoreactive neurons with classic flame-shaped NFT morphology. Threadlike immunoreactivity (panel *b*) represents neuropil threads.

CAA: cerebral amyloid angiopathy SMC: smooth muscle cell cortical tubers of tuberous sclerosis complex (39). The NFTs in this pediatric population show disorganized clumps of neurofilaments and neurotubules adjacent to cytoplasmic debris (40). The interesting question of how phospho-tau may mediate synaptic dysfunction has been addressed by several laboratories. Tai et al. (41) have shown that in AD, tau becomes hyperphosphorylated and misfolded at both presynaptic and postsynaptic nerve terminals; furthermore, accumulation of hyperphosphorylated tau oligomers (at human synaptic terminals) is associated with increased levels of ubiquitinated substrates and proteasome components, suggesting dysfunction of the ubiquitin-proteasome system (41).

CEREBRAL AMYLOID ANGIOPATHY

An often underappreciated or overlooked lesion of AD is cerebral amyloid angiopathy (CAA), sometimes described as congophilic angiopathy (42-45). CAA was the microscopic AD lesion from which Glenner & Wong (46) isolated the protein we now know as A β . CAA is less prominently featured (than SPs and NFTs) when considering AD neuropathologic lesions in part because it is variable in severity among AD patients, though when sought diligently it is identified in an estimated 85–95% of AD brains (42, 43). CAA results from a process whereby the media of parenchymal arterioles, normally composed of smooth muscle cells (SMCs), undergoes progressive loss of these SMCs and simultaneous accumulation of an eosinophilic hyaline material that has the staining properties of amyloid, that is, positivity for thioflavin S or T and congophilia, with the characteristic yellow-green birefringence (of the congophilic material) when a brain section is viewed under polarized light (34, 47). By ultrastructural examination, the amyloid has a fibrillar appearance but not the PHF morphology of NFTs. The fibrils are 7-10 nm in diameter and may be identified in close proximity to SMC debris within arteriolar media (34). It is unclear whether the amyloid fibrils are toxic to SMCs; however, tissue culture experiments suggest that mutant Aß peptides are selectively injurious to cerebral microvessel-derived SMCs (48). Hypoxia and reoxygenation of cultured brain microvessel-derived SMCs are known to mediate APP production in these cells (49). CAA may also involve cortical parenchymal capillaries and venules; at least a subset of SPs in the neocortex may be intimately associated with capillary walls and may originate from these vessels (1, 50, 51). It has been suggested that in AD, capillary A β should be distinguished from pericapillary amyloid, the former being strongly associated with the ApoE $\varepsilon 4$ isoform as a risk factor. Pericapillary A β may represent initial A β accumulation along the glia limitans, which is integral to perivascular drainage of ApoE and AB; by contrast, capillary CAA could be explained by a limited transendothelial clearance of ApoE ε 4–A β complexes (52). A shift in microglial A β binding in AD is associated with severe CAA (53).

When CAA is noted in the subarachnoid space, the amyloid deposits are usually adventitial rather than medial in the walls of larger affected arteries and have a chunky or multifocal appearance, suggesting they may have resulted from aggregates of A β in the CSF that came to lodge in arterial adventitia. CAA almost never involves the subcortical white matter, basal ganglia, brainstem, or spinal cord, but (in severe and especially familial autosomal dominant cases of AD) may involve the cerebellar molecular layer and meninges (see below) (42, 44, 47). The pathogenesis of CAA is incompletely understood and highly complex. It probably involves overproduction of A β (from APP) in or near the vessel wall, together with abnormal and/or impaired clearance of A β , probably along perivascular adventitial pathways of brain microvessels (54). Transgenic animals can be engineered to develop varying degrees of CAA, ranging from minimal to extremely severe (55).

CAA (Figure 4) is a significant cause of spontaneous nontraumatic intracerebral hemorrhage within the brains of elderly individuals—including many who do not manifest a dementing illness



Cerebral amyloid angiopathy (CAA; also known as congophilic angiopathy). (*a,b*) Micrographs from hematoxylin and eosin (H&E)-stained sections. Panel *a* shows numerous arterioles affected by CAA (*arrows*). Panel *b* shows a magnified view of an arteriole (*arrows*) in which the media (normally composed of smooth muscle cells) has been replaced by eosinophilic, slightly refractile material, with resultant thickening of the arteriolar wall. Arrowheads indicate a nearby senile plaque. (*c*) Section immunostained with anti-A β shows two arterioles with thickened walls (*arrows*), in which media is strongly immunoreactive for A β . Note the negligible parenchymal A β immunoreactivity.

or cognitive impairment at the time of their stroke; a subset of such individuals have predominantly severe CAA (with a small burden of NFTs and SPs) as their neuropathologic finding (42, 44, 47). Intraparenchymal hematomas that result from CAA are usually lobar. In some patients, multiple hematomas caused by CAA occur over months or years, with progressive neurologic decline. These substantial, invariably symptomatic, and sometimes fatal hemorrhages occur in a relatively small proportion of those with AD and CAA. CAA-related microbleeds [brain microbleeds (BMBs) detectable on high-resolution MRI scans] are now accepted as a reliable biomarker for the presence of CAA (56). The large numbers of BMBs noted by MRI are rarely, however, seen in autopsy brain specimens—a disconnect that is often the subject of spirited debate at meetings involving neuropathologists and neuroradiologists. Severe CAA has also been associated with the occurrence of cerebral microinfarcts, lesions that may worsen cognitive impairment in a patient afflicted by AD or may by themselves produce progressive neuropsychological impairment (57, 58). Some of the microfoci of encephalomalacia identified as microinfarcts at necropsy may actually be the residua of BMBs in which the blood has been resorbed. Rarely, severe CAA is associated with a prominent and rapidly progressive angiitis, sometimes described as A\beta-related angiitis (ABRA), which may have a significant granulomatous component (59). We have encountered a case in which an elderly woman presented with fatal cerebral edema, found at autopsy to be caused by ABRA (Figure 5). Is there otherwise a distinctive CAA phenotype? Arvanitakis et al. (60) have suggested that severe CAA is associated with impaired performance in perceptual speed tasks.



A β -related angiitis. Sections are from autopsy brain of a woman who had experienced rapidly progressive, fatal cerebral edema. (*a*) Low-magnification micrograph from a hematoxylin and eosin (H&E)-stained section shows numerous arterioles involved by cerebral amyloid angiopathy, many with surrounding inflammatory cells (*arrows*), including multinucleated giant cells. (*b*) Micrograph from a section immunostained with A β , showing prominent medial immunoreactivity; arrows indicate a large multinucleated (foreign body) giant cell encircling the A β .

OTHER LESIONS

Granulovacuolar degeneration (GVD, of Simchowicz) describes an abnormality in which the neuronal cytoplasm of hippocampal pyramidal cells is replaced by vacuoles containing small basophilic granules; these may represent an abnormality of neuronal autophagy. Hippocampi showing prominent GVD (**Figure 6**) also often show eosinophilic hyaline rodlike structures—rodlike bodies of Hirano—in the neuropil. GVD and Hirano bodies have been the subject of limited study and are thought to contribute minimally to AD pathogenesis and progression. Nevertheless, neurons showing GVD and Hirano bodies are a frequent finding in AD hippocampi. Ball (61, 62) has characterized the topographic distribution of both GVD and NFT neuronal change along the anteroposterior length of the hippocampus as well as within pyramidal cell layer subfields. Spongy change and reactive gliosis of varying severity are often noted in the superficial cortex (layers I–II) of AD brain; their etiology and significance are not known, but these changes need to be distinguished from the pancortical spongy change characteristic of transmissible spongiform encephalopathies (1, 2, 5).

One of the most important findings in AD brain is loss of synapses and synaptic proteins; this loss can be demonstrated biochemically or by immunohistochemistry using quantitative



Figure 6

Granulovacuolar degeneration (*arrows*) in a hippocampal pyramidal neuron from an Alzheimer's disease patient: hematoxylin and eosin (H&E)-stained section.

immunohistochemistry or a Western blot or ELISA assay (63). It was first shown dramatically in affected cortex when brain sections were immunostained with antibodies against synaptophysin (64). Quantification and subsequent interpretation of the immunohistochemical signal in such cases must be done by meticulous comparison to brain from a cognitively normal control, because the loss of synaptophysin protein may be subtle on a naked-eye inspection of the tissue section. When synaptic proteins (synaptophysin, synaptotagmin, and rab3a) were compared (by Western blot) among subjects with early- and late-onset AD and vascular dementia, the most profound loss of all synaptic proteins was noted in the early-onset AD subjects; vascular dementia brains showed minimal abnormalities (65). Soluble oligomers of A β protein impair synaptic plasticity and behavior, studied (respectively) by assessing long-term potentiation in hippocampal slice and rat experiments (66–68). An inverse relationship has also been noted between APP production (in transgenic mice) and evidence of synaptic plasticity (69, 70).

AD patients (and thus their brains) may also show evidence of comorbidity, not surprising given the many age-related diseases (e.g., cerebrovascular disease, neoplasms, infections) that may impact the aging brain (71). Coexistent Parkinson's disease changes and evidence of infarcts or hemorrhage have been seen, respectively, in as many as one-fifth and one-fourth of AD brains (72). The role of cerebrovascular disease (CVD) in contributing to cognitive impairment is described in greater detail below (73, 74).

IMMUNOHISTOCHEMICAL FEATURES

Aß protein is the major constituent of SPs (especially their cores) and CAA. The amyloid (cascade) hypothesis (more recently reworked as the amyloid oligomer hypothesis) remains a widely accepted concept of AD pathogenesis; it asserts that $A\beta$ protein deposition within the CNS is the primary cause of AD and an early critical event that promotes downstream features of the disease, such as NFT formation and synaptic injury and loss, leading to progressive dementia (1, 75, 76). It is not firmly established, however, whether aggregated fibrillar A β or soluble oligomers of the protein impair normal brain physiology. The immunohistochemical study of AD amyloid brain lesions became feasible soon after the partial peptide sequence of A β was first published (1, 2, 46); antibodies to synthetic peptides representing portions of the molecule were rapidly developed (77, 78). Numerous commercially available antibodies to $A\beta$ (various amino acid lengths), tau, ubiquitin, α -synuclein (to detect Lewy bodies) and TDP-43 are currently available to facilitate accurate immunohistochemical characterization of a given autopsy brain specimen originating from a patient with dementia. SPs are more prominently immunoreactive for the A β peptide spanning amino acids 1 to 42 (A β_{1-42}), whereas CAA is more effectively immunolabeled with antibodies to $A\beta_{1-40}$ (Figures 4, 7, and 8). Diffuse SPs are effectively shown by anti-A β antibodies, as are the amyloid cores of mature SPs. Brain specimens with abundant SPs also often show subpial accumulations of Aβ-immunoreactive material. The neuritic halos or coronas of mature SPs, NFTs, and neuropil threads are prominently immunolabeled with antibodies to phospho-tau and less prominently with anti-A β (Figures 7 and 8). In cases of severe CAA, gamma trace may also be found in affected vessel walls, and a heavily infiltrated arteriole may be surrounded by tau-immunoreactive neuritis or a halo of perivascular A β immunoreactivity (78).

STAGING ALZHEIMER'S DISEASE NEUROPATHOLOGIC CHANGE

All AD lesions described above may be encountered in the CNS of cognitively normal elderly individuals. It is therefore useful to quantify (or semiquantify) these abnormalities and assess their topographic distribution. Correlations between lesion load, severity of neuropathologic findings,



Whole-mount parallel sections of hippocampus (*left panels*) and neocortex (*right panels*) immunostained with primary antibodies as indicated in each panel. Note dense accumulations of A β (*top panels*, especially in neocortex) and phospho-tau (*bottom panels*), with sparing of subcortical white matter.

and antemortem neuropsychological testing in a given patient are important, to the extent they are feasible. The neuropathologist usually examines the brain of an end-stage subject, yet that individual may have experienced her/his maximal neurologic deficit months or years prior to death (1, 79). Correlations between neuropathologic autopsy findings and neuroimaging or neuropsychology are likewise problematic because the patient may not have been well enough to be tested for many months or years before passing. Rarely, biopsies are carried out on AD subjects—for example, incidental sampling of the brain during therapeutic insertion of a ventricular catheter. Small studies have used biopsy and autopsy data from the same patient to describe the progression of AD lesions over years (80). These studies have proven that there are significant AD lesions in the brain of an individual who is at a cognitively early stage of clinical symptoms.

Attempts have been made, since the 1980s, to standardize neuropathologic diagnostic criteria for AD/SDAT. A consensus conference in the 1980s resulted in the widely used Khachaturian criteria for the neuropathologic diagnosis of AD—criteria that utilized nonimmunohistochemical approaches for evaluation of the brain (81). These were modified and updated by the Consortium to Establish a Registry for Alzheimer's Disease (CERAD) (9, 72), a highly productive group effort to simplify AD diagnosis with easy-to-use templates and diagrams. The Braak criteria for AD severity (25, 26) assume a progression of neuropathologic abnormalities (predominantly NFT and neuropil thread accumulation) from the transentorhinal cortex (stages I and II) to the hippocampus (III and IV), finally with widespread involvement of the neocortex (stages V and VI).

The National Institute on Aging (NIA)-Reagan Institute criteria for the neuropathologic diagnosis of AD came into widespread use in the late 1990s and have been tested and operationalized



Sections of hippocampus immunostained with three different primary antibodies: $A\beta_{1-40}$, $A\beta_{1-42}$, and phospho–paired helical filament–tau (phospho–PHF-tau). Boxes in each of the upper three panels indicate the region magnified in the corresponding lower panel. Note prominent senile plaques (demonstrated most effectively with anti- $A\beta_{1-42}$); tau immunostaining highlights immunoreactive neurons in the granule cell layer.

(82). These criteria assign a high, intermediate, or low likelihood that a given individual's dementia was due to AD neuropathologic features. One such study found a good correlation between a high NIA-Reagan probability likelihood of AD and clinical dementia but further ascertained that the older Khachaturian and CERAD criteria correlated fairly well with those of NIA-Reagan (82). Occasional cases arise—especially among the oldest old—in which advanced AD neuropathologic changes were clearly present in the brain of a subject who was known to be cognitively intact until shortly before death (31), though such rare cases may be outliers. Quantification of AD neuropathologic change is increasingly facilitated by the ability to digitize immunostained glass slides, retain the images as a permanent electronic record of a given autopsy, and use these digital images as a starting point for further quantitative morphometric analysis. A novel antibody capture assay has recently been developed for immunostained paraffin sections, facilitating quantitative antigen assays in such tissue sections (83).

In recent years, new guidelines for the neuropathologic assessment of AD have been developed (84, 85). These have built on the Braak, NIA-Reagan, and CERAD approaches and recognize that the neuropathologic changes of AD may occur in the apparent absence of cognitive impairment. They are based upon an assessment of three spheres of neuropathologic change, including (*a*) the extent of A β deposition [using the scoring system of Thal et al. (33)], (*b*) staging of NFT burden (collapsing the six Braak stages into a more simplified three), and (*c*) assessing density of neuritic SPs, as was initially proposed by CERAD. Furthermore, these NIA–Alzheimer's Association guidelines provide more detailed and specific approaches to evaluating comorbid conditions such as Lewy body disease, vascular brain injury, and hippocampal sclerosis. The final AD score

is thus formulated so as to allow the tissue evaluator to state that a given case does not show AD pathologic change or that it has a low, intermediate, or high likelihood of AD. This system, while recently developed and published, is currently being evaluated for interobserver and intercenter reproducibility.

MILD COGNITIVE IMPAIRMENT

MCI emerged as a concept and nosologic entity in the mid- to late 1990s. It describes subclinical complaints of memory dysfunction (usually in elderly subjects). Deficits in some cognitive tests have been noted in MCI subjects, but normal intellectual function is generally preserved (86). Affected individuals have a high probability of evolving towards AD/SDAT. MCI subjects are considered a potentially important group to study in terms of early intervention, that is, at a time prior to irreversible brain changes, when their cognitive deficit is relatively mild and might be stabilized or improved. Neuropathologic studies on such patients are infrequent, because (unlike end-stage AD subjects) they are not medically unwell and prone to life-threatening events. However, when autopsies have been performed on individuals with amnestic MCI (aMCI), the neuropathologic findings suggest a transitional state of evolving AD. Braak stage III and IV AD neuropathologic change was most commonly encountered, but two (of fifteen) patients in one study had neocortical pathologic abnormalities (Braak stage V). Over half of the patients who came to necropsy had evidence of significant CVD, including hippocampal injury that was often interpreted as ischemic in origin, that is, hippocampal sclerosis (87, 88). The concept of aMCI has been present for a sufficiently long time that many affected patients have progressed to dementia. Neuropathologic studies of these subjects show that approximately two-thirds had AD changes at necropsy, and the remainder showed heterogeneous neuropathologic abnormalities that explained their cognitive decline, including FTLDs, hippocampal sclerosis, and nonspecific tauopathy (89). Data from the Religious Orders Study show that among over 130 individuals with MCI who underwent autopsy, 54.5% had pathologically confirmed AD and 19.4% had mixed pathologies, usually including ischemic infarcts or Lewy bodies (90). As in the Mayo Clinic study (87), therefore, CVD was a common accompaniment of AD—indeed, macroscopic brain infarcts (in the absence of pathologically confirmed AD) accounted for 13.3% of aMCI cases and 18.6% of nonamnestic MCI cases.

FAMILIAL ALZHEIMER'S DISEASE

AD is infrequently inherited as an autosomal dominant trait, the result of mutations in one of three genes: the amyloid precursor protein (*APP*) gene on chromosome 21, presenilin 1 (*PSEN1*) on chromosome 14, and presenilin 2 (*PSEN2*) on chromosome 1. *PSEN1* mutations are the most common, whereas *PSEN2* mutations are very rare (1, 2). Disease onset is variable but often occurs at an early age (in the third or fourth decade of life). Down syndrome (DS, trisomy 21) subjects will invariably develop AD neuropathologic change if they survive to the fourth or fifth decade. (Note that the ApoE ε 4 isoform is a genetic risk factor for developing AD, but is not in itself a disease-determining abnormality.) Autosomal dominant variants of AD are of obvious interest to investigators; a subject with an *APP*, *PSEN1*, or *PSEN2* mutation will invariably develop the disease; thus, neuropsychological and neuroimaging features of reliable disease progression, as well as other putative CSF biomarkers, can be studied in an afflicted individual, even at an early age. Such subjects are also of great interest in examining prevention and treatment strategies that target early stages of the disease. *APP* mutations are found in and adjacent to the short A β -coding domain of the gene, flanking the β - and γ -secretase sites at which A β is cleaved from APP (91).



Section of cerebellum from a familial (autosomal dominant) Alzheimer's disease subject with a *PSEN1* mutation. Note prominently $A\beta$ -immunoreactive deposits throughout the molecular layer (*arrowbeads*). Arrows indicate prominent cerebral amyloid angiopathy affecting many leptomeningeal arterioles.

Although most *APP* mutations cause AD, an Icelandic group has discovered a coding mutation in this gene (A673T) that protects against AD cognitive decline—apparently as the result of an approximately 40% reduction in the formation of amyloidogenic peptides (92). Autopsy data on this unique cohort of protected subjects have not been published. All *APP* and *PSEN* mutations are thought to mediate their effects through their impact on A β processing from APP (93). Of interest, *APP* mutations in any of three adjacent codons—692, 693, and 694—result in a phenotype dominated by severe CAA; in the hereditary Dutch form of CAA, the neuropathologic features are almost exclusively vascular, including numerous infarcts and hemorrhages throughout the brain in the presence of some SPs but in the absence of significant tau pathology (47, 56). Quite severe CAA may also be encountered in the brains of *PSEN1* mutation carriers when the mutation is beyond codon 200 (unlike in *APP*, *PSEN1* mutations are found throughout the gene, which encodes a multi-membrane-spanning protein). We have found rather prominent A β deposition in the cerebella of some familial AD patients, in the form of both SPs and CAA (**Figure 9**).

There is increasing interest in genes that may modify expression of the AD phenotype, analogous to the effect of ApoE ε 4. Many of these have been discovered using the powerful technology of genome-wide association studies (for review, see 94). These include complement component (3b/4b) receptor 1 (CR1), on chromosome 1q32; bridging integrator 1 (BIN1), on chromosome 2q14; phosphatidylinositol-binding clathrin assembly protein (PICALM), on chromosome 11q14; membrane-spanning 4-domains, subfamily A, members 6A and 4E (MS4A6A and MS4A4E), on chromosome 11q12.1; ATP-binding cassette, subfamily A, member 7 (ABCA7), on chromosome 19p13.3; and clusterin (CLU), on chromosome 8p21-p12. Examining the neuropathologic effects (if any) of these genes (and polymorphisms and amino acid changes within them) will be a challenge to neuropathologists. As Holton et al. (94) have stated, "Measuring changes in damage induced expression in tissue with changing cell populations and developing rigorous algorithms to interpret such data is problematic." One of the most interesting observations has been that variants in triggering receptor expressed on myeloid cells 2 (TREM2) result in susceptibility to late-onset AD (95, 96, 97). Though the significance of this association remains the subject of debate, it suggests a plausible mechanism of disease pathogenesis: TREM2 is an innate immune receptor expressed on the membranes of some myeloid cells (including tissue macrophages and brain microglia). TREM2 on microglia may be critical to the clearance of neural debris of the lesioned CNS (95)though the specific research questions to be addressed will need to be more clearly focused, given that the ligand of the lesioned neural tissue recognized by TREM2 is as yet unknown.

CLINICOPATHOLOGIC CORRELATION IN DEMENTED SUBJECTS

Clinicopathologic analysis of subjects with dementia was, until the refinement of structural and metabolic antemortem brain imaging, the definitive way to link structural brain lesions with neurobehavioral and neurologic brain abnormalities. Despite their seminal contributions to understanding neurodegenerative diseases, autopsy studies are frequently criticized for their shortcomings. They often focus on studying highly selected patients who are not necessarily representative of a given population (1, 2, 5). They examine tissue at the end stage of a disease process that began years—possibly decades—prior to the affected subject's demise; therefore, causal or precipitating events that are of etiologic importance in AD/SDAT may be difficult or impossible to decipher. As one recent multicenter study has affirmed (24), "Studies on human subjects with autopsy confirmation entail numerous potential biases that affect both their general applicability and the validity of the [clinicopathologic] correlations." However, its well-argued conclusion was that AD is indeed a specific disease entity, one defined by the presence within brain of A β SPs and tau-immunoreactive NFTs. It further concluded that although A β SPs may play a key role in disease pathogenesis, cognitive decline and impairment correlate best with the burden of neocortical NFTs.

A recent theme of autopsy studies of dementing disorders has been the high frequency of non-AD comorbidity. In a large Austrian autopsy study and review of the literature (98), AD was the most frequent neuropathologic diagnosis (in >40% of subjects), but AD together with significant CVD was found in approximately 25% of subjects. Pure vascular dementia was identified in 10% of subjects and AD along with diffuse Lewy body disease in a similar percentage (these are described as mixed dementias). Cerebrovascular pathology was subclassified into four degrees of severity, depending upon the frequency and extent of lacunes, cystic infarcts and microinfarcts or hemorrhages, CAA, and hippocampal injury. The brains of nonagenarians appear to be impacted less by AD than by other comorbid conditions, especially sequelae of CVD (7, 24). Neuropathologic study of 108 decedents in the 90+ Study (based in Southern California) found that in 66 subjects with clinical AD-type dementia, quantitative immunohistochemistry showed a strong correlation between cognitive impairment and "area of cortex occupied" by $A\beta$ and tau immunoreactivity; Braak stage also correlated with dementia, whereas semiquantitative SP scores did not. The 5.6% estimated area of cortex showing Aß immunoreactivity in those without dementia nearly doubled, to 10.7%, for demented subjects (99). Comparable figures for tau immunoreactivity within hippocampus were 4.6% (nondemented) and 7.2% (demented). Hippocampal sclerosis was noted in 42% of subjects, and this correlated strongly with neuronal TDP-43 immunoreactivity.

Findings from the Adult Changes of Thought study (Seattle) as well as the Religious Orders Study and the Memory and Aging Project (Rush University, Chicago) highlight the etiologic complexity of dementing syndromes in older populations (100). Cerebral microinfarcts appear to be an especially important neuropathologic correlate of cognitive decline or dementia resulting from vascular brain injury in population-based studies. Review of autopsy data from the UCLA Alzheimer Center neuropathology database shows an age-related increase in the frequency of cerebral microinfarcts, ranging from less than 5% of brains examined from those ≤ 60 years of age to over 35% in those >90 years of age. Schneider et al. (101, 102) have observed that community-based subjects carrying the clinical diagnosis of AD had less severe AD neuropathologic change and more ischemic brain injury than clinic-based patients seen at the Rush Alzheimer Center. Clinic-based individuals. The issue of whether and how CVD causes or contributes significantly to vascular cognitive impairment (a pre–vascular dementia condition analogous to MCI as pre-AD) has been addressed in the proceedings of a multidisciplinary workshop (103).

Table 1 Biomarkers of potential value in supporting the clinical diagnosis of Alzheimer's disease/senile dementia of Alzheimer type

Method	Measures
Thorough neurologic and neuropsychological examination of the subject	Emphasize mental status, memory storage and retrieval, focal signs
Neuroimaging (structural, magnetic resonance imaging)	Ventricular enlargement (ventriculomegaly) Thinning of the cortical ribbon Hippocampal atrophy/enlargement of the temporal horn of the lateral ventricle Brian microbleeds (for cerebral amyloid angiopathy)
Neuroimaging (metabolic)	Pittsburgh compound B (PiB), labeled with ¹⁸ F or ¹¹ C: florbetapir or amyvid [¹⁸ F]Fluoroethyl-methyl-amino-2-naphthyl-ethylidene malononitrile (FDDNP) [¹⁸ F]Fluoro-dexoyglucose positron emission tomography Tau markers
Cerebrospinal fluid testing	Amyloid beta 1–42 Total tau and phospho-tau 14-3-3 protein (to rule out spongiform encephalopathy)
Blood testing	Apolipoprotein E isoforms (polymerase chain reaction–based assay) Neuron-specific enolase, S100B (as a measure of brain injury)

BIOMARKERS OF ALZHEIMER'S DISEASE/SENILE DEMENTIA OF THE ALZHEIMER TYPE AND OTHER DEMENTIAS

Biomarkers of various human disorders (especially neurodegenerative diseases) have become a major theme of twenty-first century biomedical research. Biomarkers serve as proxies or surrogates for specific pathophysiological disease processes (104, 105). Identifying quantifiable markers of disease—whether by neuroimaging, CSF, or blood markers—has the potential (*a*) to allow for early identification of possibly presymptomatic subjects at risk for developing AD and (*b*) to provide measures by which disease severity and progression may be evaluated; thus, such biomarkers may be invaluable in providing endpoints (or objective milestones) for clinical therapeutic trials. Careful and detailed physical examination of a human patient thus in itself represents a type of biomarker. **Table 1** summarizes the major biomarkers of importance in individuals suspected of having AD or another neurodegenerative disease. A role for the neuropathologist is to provide validation of biomarkers by providing feedback on putative structural (autopsy) brain changes that a given biomarker is thought to measure or reflect (1, 6). Of the large numbers of AD and MCI subjects who have had structural and/or metabolic brain imaging, for example, only a small percentage have had confirmatory neuropathologic brain examination, frequently with a considerable interval between the radiographic study and necropsy.

As the Alliance for Aging Research AD Biomarkers Work Group has stated, there are five important and established AD biomarkers: findings from three modes of neuroimaging [amyloid PET, fluorodeoxyglucose (FDG)-PET, and structural MRI] (**Figure 10**) and levels of two CSF proteins [A β and tau, including total tau (t-tau) and phospho-tau (p-tau)]. MRI images can be evaluated using visual assessment, quantitative region of interest–based techniques, automated and semiautomated methods, and quantitative voxel-based methodology (104). Studies suggest that the cortical degeneration in AD is the reverse of the normal developmental sequence within brain, a process described as retrogenesis (6). Both cross-sectional and longitudinal study approaches have been used with all neuroimaging techniques. Global atrophy can be quantified using the technique of boundary shift integral or tensor-based morphometry. Correlative MRIneuropathologic studies show that quantitative measures of brain volume loss correlate well with



Structural (MRI) and amyloid PET imaging of axial slices of brain from a control subject (*two columns at left*) and an Alzheimer's disease (AD) subject (*two columns at right*). Note the prominent Pittsburgh compound B (PiB) signal in the AD brain (*far right column*). Images courtesy of William Jagust, University of California, Berkeley.

pathologic measures of the severity of neurodegeneration. Regionally accentuated atrophy (e.g., affecting the mesial temporal lobe) is especially characteristic of AD (105). Volumetric MRI scans (emphasizing region-specific atrophy) are also useful in distinguishing AD from frontotemporal dementia (106, 107). Ventricular volume increases over time may also be a marker for AD and vascular brain injury in elderly individuals (108).

Brain A β can be imaged in vivo using Pittsburgh compound B (PiB) labeled with ¹¹C ([¹¹C]PiB) or ¹⁸F ([¹⁸F]PiB) (109). PET scanning using fluorodeoxyglucose (FDG-PET) provides a measure of brain glucose utilization and thus of cortical metabolism. Of interest, whereas structural (MRI) imaging has emphasized mesial temporal (e.g., hippocampal) atrophy (see above), FDG-PET scanning shows hypometabolism in the lateral temporoparietal and medial parietal cortex, including the precuneus and posterior cingulate gyrus (3, 105). It is now possible to study a living brain using all of these modalities and to integrate or coregister the resultant data and correlate them with CSF markers (110–114). Much of this work has been accomplished through the Alzheimer Disease Neuroimaging Initiative (ADNI), a multicenter, multidisciplinary investigation charged with studying large numbers of subjects with early AD or MCI and comparing them with normal controls (110). PiB scanning has been performed on cognitively normal elderly subjects, who were also assessed for evidence of gray matter atrophy (115). Reading of PiB scans often dichotomizes subjects as being PiB⁺ or PiB⁻. Older subjects showed age-related gray matter atrophy throughout the CNS, regardless of A β deposition. Among PiB⁺ subjects, amyloid burden was associated with gray matter atrophy within frontal, parietal, and temporal cortices. Neuropathologic examination of a PiB⁺

PiB: Pittsburgh compound B

and a PiB⁻ case (postmortem examination 10 and 17 months postimaging) showed that A β amyloid pathology within brain may be associated with undetectable or low levels of [¹¹C]PiB retention (116, 117). A considerably larger study of [¹⁸F]PiB (florbetapir), correlating antemortem imaging with autopsy findings, showed the sensitivity and specificity of florbetapir to be 92% and 100%, respectively, among subjects who had been scanned within 2 years prior to death and 96% and 100%, respectively, among those who had autopsy within 1 year (118). In autopsy brain tissue, PiB appears to bind to SPs immunoreactive for either A β_{1-42} or A β_{1-40} and to microvascular amyloid. Progressive hippocampal atrophy can be evaluated by measuring changes in hippocampal volume over time (119). Antemortem imaging of CNS phospho-tau appears to be more difficult, though some studies suggest it can be done using [¹⁸F]fluoroethyl-methyl-amino-2-naphthyl-ethylidene malononitrile (FDDNP); studies using this ligand have been performed in subjects with MCI (13).

Within the CSF, levels of t-tau, p-tau, and β -protein (usually A β_{1-42}) can be measured, though longitudinal study of these peptides may be challenging because (potentially unpleasant) serial lumbar punctures are required (104, 120, 121). A clinician attempting to differentiate a patient with FTLD from one with AD/SDAT can do so using the CSF t-tau:A β ratio; A β levels decrease with AD/SDAT progression, whereas t-tau levels tend to increase. These levels and ratios may also be helpful in predicting which MCI subjects will progress to AD. There appears to be an inverse correlation between CSF A β_{1-42} levels and SP pathologic change (at autopsy), suggesting sequestration of this peptide into extracellular SPs (120). Measures of premorbid cognitive reserve (level of education, occupation, premorbid IQ) may also be reflected in the CSF levels of these peptides (119). CSF A β and amyloid PET measurements of peptide were shown to be consistent, with a small but significant number of discordant subjects, in a large ADNI study (122). A major challenge in future studies of CSF biomarkers will be identifying reliable markers of FTLD subtypes, as well as AD cases with significant comorbidity (e.g., cerebral microinfarcts) that are currently suboptimally identified on neuroimaging studies.

Analysis of blood samples is less likely to be informative in providing biological insights into AD/SDAT, though it may be of some limited value. In a clinical setting, blood tests are valuable in providing important information in a given patient on metabolic disease that may mimic a neurodegenerative disorder. ApoE genotype is easily assessed using a polymerase chain reaction–based assay on a blood sample. One study has concluded that low plasma $A\beta_{1-42}:A\beta_{1-40}$ ratios may be associated with increased imminent risk of developing MCI and AD/SDAT (123). Other studies suggest some clinical or prognostic value in measuring blood levels of oxysterols, isoprostane, and plasma signaling proteins (124, 125). Plasma A β levels appear not to reflect brain A β levels as measured at necropsy by immunohistochemistry or biochemical assays (126).

EFFECTS OF TREATMENT ON ALZHEIMER'S DISEASE NEUROPATHOLOGIC FINDINGS

Neuropathologic studies are likely to be crucial in documenting how and why (yet to be developed) treatments for AD/SDAT work or fail to work. AD is currently neither treatable nor preventable. Cholinesterase inhibitors or memantine provides temporary symptomatic improvement in some patients (75). In the late 1990s and early 2000s, based upon success in clearing brain A β from transgenic mice using an amyloid vaccine, a clinical vaccine trial was initiated in humans. Unfortunately, 5–6% of immunized subjects developed a rapidly fatal meningoencephalitis. Autopsy studies on a few of these subjects showed remarkable findings. There was evidence of A β clearance from cerebral cortex, though NFT and neuritic changes remained, as did CAA (127). The meningoencephalitis appeared to be mediated by T lymphocytes. A β immunoreactivity was associated

with (possibly activated) microglia, and the subcortical white matter showed extensive infiltration by macrophages. A second case report (128) described similar findings in an immunized subject and commented on the presence within cerebral cortex of dense SPs surrounded by microglia and multinucleated giant cells. T lymphocytes present within brain included predominantly CD8⁺, CD4⁺, CD3⁺, and CD5⁺ cells; B cells and cytotoxic T cells were absent. Multiple cortical BMBs (usually a marker for severe CAA) were observed. These fascinating initial case reports presented data on a very limited number and subset of patients and made inferences of dynamic events pertinent to A β clearance that could not be compared with appropriate controls or prior tissue examination, as no patients had undergone brain biopsies. Much larger numbers of subjects have now been studied.

There has been subsequent detailed clinical, neuroimaging, and neuropathologic analysis of many vaccine-treated subjects, and observations in these individuals have been compared with those in nonvaccinated controls (129). Though the *n* of subjects in these studies has been comparatively small, several intriguing observations have emerged. Mean A β load was lower in the immunized group than in age-matched untreated controls. The degree of SP removal varied with mean antibody response attained. Dementia did not improve in treated subjects, nor did overall survival. A β clearance has been documented in other studies (130), as has diminution of neuritic change in SPs, though NFTs, neuropil threads, and CAA persisted. No evidence for a beneficial effect on synapses and no morphologic evidence of synaptic integrity were found (131).

Boche et al. (131, 132) and Weller et al. (54) have hypothesized that SP A β is solubilized by antibodies generated during immunization and that the peptide then drains along perivascular pathways in the CNS, detectable (by immunohistochemistry) as severe CAA. When immunized subjects were compared with controls, the former had 14 times as many $A\beta$ -containing blood vessels within cortex and 7 times as many within the leptomeninges; the parenchymal CAA was severe, involving the full thickness of many arteriolar walls. Significantly more $A\beta_{1-40}$ was seen in the immunized cases, which also showed a higher density of BMBs and microvascular lesions. An intriguing observation was that two of the longest survivors (4 and 5 years after initial immunization) had virtually complete absence of SPs and CAA within their brains. Autopsy studies on the effect of a given treatment are limited by the fact that the treated brain is examined at only one point in time, as the authors of these papers acknowledge-this and the small number of specimens available for analysis limit detailed biochemical study (133). However, imaging of brains using amyloid PET (e.g., with PiB) will likely allow for dynamic observations of A β deposition and clearance over time—amyloid PET imaging can be carried out over several time points—but autopsy examination of the brain will remain crucial. Comparisons of treated versus untreated subjects (rigorously matched for age, disease duration, etc.) will always be valuable. Recently, a detailed analysis of inflammatory components within autopsy brain of Aß immunized versus nonimmunized subjects has been undertaken (134). Quantitative analysis was undertaken of various microglial and inflammatory markers, including CD68, macrophage scavenger receptor A, CD64, CD32, Iba-1+, C1q, the T cell marker CD3, and IgG. Levels of CD68, macrophage scavenger receptor A, CD64, and CD32 were significantly lower in immunized than nonimmunized subjects, though there was no significant difference in Iba-1+ load, numbers of Iba-1+-positive cells, IgG load, or numbers of T cells. In this study, there was evidence of substantial reduction of A β_{1-42} load (by over 80%) and phospho-tau (by 40%), though the tau decrease appeared to impact mainly neuronal processes rather than NFTs. This paper provides substantial evidence for the importance of microglial function or activation (as a function of vaccination) in phospho-tau accumulation or clearance within the brain (see also the comments on TREM2 above).

Recent high-profile reports have presented largely negative results of AD immunotherapy using humanized anti-A β antibodies, bapineuzumab and solanezumab (76, 135, 136). Clinical and neuropsychological outcomes did not improve in treated patients, despite significant improvements in CSF phospho-tau levels in ApoE ε 4 carriers. Decreased rate of A β accumulation within brain was seen (on PiB PET scans) in ApoE ε 4 carriers. A significant complication of both humanized antibodies was cerebral edema or hemorrhage, possibly mediated by the impact of the immunotherapy on CAA (see the discussion above). Bapineuzumab significantly alters A β composition, according to neuropathologic and biochemical analysis of a small number of specimens (133). It has also been shown to neutralize the detrimental effects of A β oligomers on synaptic plasticity in an animal model (137).

The search for an effective candidate AD therapy continues. The use of A β -degrading enzymes (e.g., neprilysin, cathepsin B) has been suggested as one treatment strategy (138). However, an approach to preventing the extensive tau pathologic change that characterizes AD brain will also need to be developed, and indeed may be a key to definitively improving cognitive function.

CEREBROVASCULAR DISEASE AND ALZHEIMER'S DISEASE/SENILE DEMENTIA OF THE ALZHEIMER TYPE

Just as the aging human brain is at increased risk, with every decade, for the development of AD/SDAT, so is it prone to manifest the consequences of CVD. In considering CVD as a cause of cognitive impairment, there are several important themes: (a) pure CVD (with minimal AD brain changes) as a cause of cognitive impairment (ischemic vascular dementia or IVD) (74, 139–142); (b) comorbidity between AD and CVD in a given brain (sometimes described as mixed dementia), leading to more severe impairment than would result from AD changes alone (73, 143–147); (c) the hypothesis that cerebral atherosclerosis itself is a major risk factor for AD/SDAT; and (d) blood-brain barrier dysfunction as a key contributing factor to AD pathogenesis and progression. I emphasize above that a microvascular lesion commonly found in the brains of aged individuals (and more severe in those with AD/SDAT), CAA, is associated with brain parenchymal abnormalities including large hematomas, BMBs, and microinfarcts (42, 58, 148). Brains of the elderly and those with AD also show a significant degree of arteriolosclerosis, sometimes described as lipohyalinosis, and these microangiopathies may synergistically contribute to AD progression (149, 150). In large autopsy series, with the inherent bias of such investigations, relatively pure IVD accounts for 7-10% of cases, genuinely mixed findings (significant CVD and AD neuropathologic changes) for 3-5% of cases, and AD with a minor CVD component for 20–40% of cases; estimates vary widely depending upon the center reporting them (98, 146). Neuropathologic substrates of IVD include cystic infarcts, usually in the territories of large arteries or their branches, lacunar infarcts (grossly visible in the autopsy brain or on neuroimaging) smaller than 1.0 cm in greatest dimension, and microinfarcts (not visible grossly or on neuroimaging but seen in histologic sections of the brain; see the discussion above) (74, 149–152). Cystic infarcts in certain regions are especially likely to cause memory storage and retrieval deficits as well as cognitive impairment; these regions include the hippocampi and angular gyrus. Ischemic changes within hippocampi (resembling hippocampal sclerosis that causes temporal lobe epilepsy)-especially common in the oldest old-are an important substrate for neurobehavioral changes that may resemble AD/SDAT, with prominent deficits in memory storage and retrieval (153–156). Because most neuropathologic examinations of the brain entail sampling a relatively restricted region of the hippocampus, neuropathologic change within it (including linear scars and small infarcts) is almost certainly underestimated and only becomes apparent when serial blocks of this structure are carefully examined, sometimes using morphometric techniques (157). Though it was at one time suggested that AD/SDAT is a hippocampal dementia, the prevailing view is now that neocortical lesions are a necessary substrate of cognitive decline (158).

Abnormalities of cerebral microcirculation (i.e., the neurovascular unit, which includes the capillary and arteriolar endothelium and surrounding pericytes, SMCs, and astrocytes) have been suggested as key elements in AD/SDAT pathogenesis (159-161). The role of the perivascular spaces (surrounding brain parenchymal vessels) in clearing brain parenchymal A β is discussed above, including with reference to the consequences of A β vaccination. A β transport through the blood-brain barrier endothelium is mediated by two main receptors, the low-density lipoprotein receptor-related protein (LRP1) and the receptor for advanced glycation end products (RAGE) (159, 160). Subtle abnormalities of the blood-brain barrier suggestive of leakiness have been demonstrated by ultrastructural studies (162). Whether large artery atherosclerosis (e.g., affecting branches of the circle of Willis) contributes directly to AD/SDAT pathogenesis has been the subject of lively and as yet unresolved debate. Most of the studies supporting or questioning such an association have been based upon autopsy data (163–167). The Baltimore Longitudinal Study of Aging, for example, concluded that atherosclerosis is an important and independent risk factor for dementia, but one unrelated to AD/SDAT pathologic change (167). Data from our longitudinal California-wide study of vascular factors important in brain aging suggest a similar conclusion (168). We have also found an interesting association of cerebral atherosclerosis with brain cystic infarcts (well known previously) and CNS microinfarcts (169). Others, however, report a compelling association between circle of Willis atherosclerosis and SPs and NFTs within the brain (163). One problem with such investigations is that large cervical arteries—atherosclerosis within which significantly impacts brain parenchyma, arguably more prominently than circle of Willis atheroma-are almost never examined in autopsies.

CONCLUDING REMARKS AND FUTURE DIRECTIONS

This article has intentionally focused on selected clinicopathologic aspects of AD/SDAT, while avoiding others. The contributions of transgenic animal models to understanding AD pathogenesis have been immense but are beyond the scope of this review (55, 170). The amyloid (cascade) hypothesis has been the foundation upon which much modern AD research is based-yet strategies that appear to be effective in removing this protein from the brain have led to negligible clinical improvement in treated subjects and to only modest decreases in phospho-tau abnormalities within treated brains (see the section Effects of Treatment on Alzheimer's Disease Neuropathologic Findings, above). Several excellent recent reviews have discussed practical and theoretical frameworks for considering AD/SDAT in the context of brain aging and important genetic abnormalities that influence this process (171). Jagust (3, 105) has stressed the neuroanatomical heterogeneity of brain aging, including varying degrees of dysfunction in its different regions—for example, the medial temporal lobe memory system and a frontostriatal executive system. Huang & Mucke (172) emphasized that AD probably has multiple causes with complex interactions, thus concluding that investigative and drug discovery and development efforts should be diversified to realistically address the multifactorial nature of the disease; monotherapy is extremely unlikely to work. The emphasis of much research is on expanding from the level of synapses, where (as discussed above) AD produces significant abnormalities, to the level of neural networks (4). Skeptics of the amyloid (cascade) hypothesis abound and provide interesting, sometimes controversial points of view.

Ball et al. (173) have proposed a role for viral (herpes simplex) infection in intracerebral propagation of AD pathologic change. It has also been hypothesized that A β may seed and self-propagate by a mechanism analogous to that seen in prion diseases; indeed, transsynaptic spread of tau and α -synuclein may be an important mechanism that explains stereotypical patterns of lesion spread within the brain (174). Stohr et al. (175) have demonstrated the ability of A β aggregates to selfpropagate and spread throughout the brains of transgenic mice after intracerebral inoculation; they were able to show this propagation using either purified A β aggregates derived from brain tissue or aggregates composed of synthetic A β . They have taken the bold (but possibly justified) step of referring to $A\beta$ as $A\beta$ prions (175). Hamaguchi et al. (176) have shown that the presence of A β seeds, and not the age of the host animal being tested, is crucial to the initiation of A β aggregation within the CNS and leads to spread of the protein throughout the brain. Transsynaptic progression of A\beta-induced neuronal dysfunction has been demonstrated within the entorhinalhippocampal network (177). Analogous phenomena have been suggested or demonstrated for tau within both human and mouse brains. Braak & Del Tredici (178) have made the provocative observation that tau abnormalities may be seen in the brains of individuals younger than 20 years of age-for example, in the pontine locus ceruleus (which projects to cortex)-supporting the idea of neuron-to-neuron propagation of abnormal cytoskeletal protein. Injection of brain extract from mutant P301S tau-expressing mice into the brains of transgenic wild-type tau-expressing animals induced assembly of wild-type human tau into filaments and the spread of pathologic lesions from the injection site(s) to nearby brain regions (179). Conversely, Perry and colleagues (180, 181) have repeatedly suggested that the microscopic lesions associated with AD are a suboptimal, even poor, therapeutic target, and that AD pathology is indicative of an active host response or environmental adaptation to pathophysiologic mechanisms such as oxidative stress within the CNS.

The full neuropathologic characterization of evolving and unforeseen types of neurodegenerative disease will provide stimulating full employment for neuropathologists in the decades to come (182–186). They will be charged not only with characterizing abnormal shadows and signals detected by neuroradiologists, but also with providing feedback to clinicians on how well therapies aimed at clearing abnormal proteins from the brain have worked (or failed to work). Finally, will removing abnormal brain proteins that accumulate in neurodegenerative diseases lead to clinical improvement in patients? This will be the crucible in which novel therapeutic strategies will need to be tested; neuropathologic evidence will be crucial to interpreting the results of new and innovative therapies.

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LITERATURE CITED

- Dickson DW, Weller RO, eds. 2011. Neurodegeneration: The Molecular Pathology of Dementia and Movement Disorders. Oxford, UK: Wiley-Blackwell. 2nd ed.
- Ellison D, Love S, Chimelli L, Harding BN, Lowe JS, et al. 2013. Neuropathology: A Reference Text of CNS Pathology. Edinburgh/London: Mosby. 3rd ed.
- Jagust W. 2013. Vulnerable neural systems and the borderland of brain aging and neurodegeneration. Neuron 77:219–34
- Palop JJ, Mucke L. 2010. Amyloid-β-induced neuronal dysfunction in Alzheimer's disease: from synapses toward neural networks. *Nat. Neurosci.* 13:812–18

- Vinters HV, Farrell MA, Mischel PS, Anders KH. 1998. Neurodegenerative diseases. In *Diagnostic Neuropathology*, pp. 453–507. New York: Marcel Dekker
- Thompson PM, Vinters HV. 2012. Pathologic lesions in neurodegenerative diseases. *Mol. Biol. Transl. Sci.* 107:1–40
- Nelson PT, Head E, Schmitt FA, Davis PR, Neltner JH, et al. 2011. Alzheimer's disease is not "brain aging": neuropathological, genetic, and epidemiological human studies. *Acta Neuropathol.* 121:571–87
- Kawas C, Gray S, Brookmeyer R, Fozard J, Zonderman A. 2000. Age-specific incidence rates of Alzheimer's disease: the Baltimore Longitudinal Study of Aging. *Neurology* 54:2072–77
- Mirra SS, Heyman A, McKeel D, Sumi SM, Crain BJ, et al. 1991. The Consortium to Establish a Registry for Alzheimer's Disease (CERAD). Part II. Standardization of the neuropathologic assessment of Alzheimer's disease. *Neurology* 41:479–86
- Jellinger KA. 2002. Alzheimer disease and cerebrovascular pathology: an update. *J. Neural Transm.* 109:813–36
- 11. Rahimi F, Bitan G, ed. 2012. Non-Fibrillar Amyloidogenic Protein Assemblies—Common Cytotoxins Underlying Degenerative Diseases. Dordrecht, Neth.: Springer
- Mintun MA, LaRossa GN, Sheline YI, Dence CS, Lee SY. 2006. [¹¹C]PIB in a nondemented population: potential antecedent marker of Alzheimer disease. *Neurology* 67:446–52
- Small GW, Kepe V, Ercoli LM, Siddarth P, Bookheimer SY, et al. 2006. PET of brain amyloid and tau in mild cognitive impairment. N. Engl. J. Med. 355:2652–63
- Small GW, Komo S, La Rue A, Saxena S, Phelps ME. 1996. Early detection of Alzheimer's disease by combining apolipoprotein E and neuroimaging. *Ann. N.Y. Acad. Sci.* 802:70–78
- 15. Vinters HV. 2007. Imaging cerebral microvascular amyloid. Ann. Neurol. 62:209-12
- 16. Goedert M, Ghetti B. 2007. Alois Alzheimer: his life and times. Brain Pathol. 17:57-62
- Kovacs GG, Botond G, Budka H. 2010. Protein coding of neurodegenerative dementias: the neuropathological basis of biomarker diagnostics. *Acta Neuropathol.* 119:389–408
- 18. Querfurth HW, LaFerla FM. 2010. Alzheimer's disease. N. Engl. J. Med. 362:329-44
- Armstrong RA, Ellis W, Hamilton RL, Mackenzie IRA, Hedreen J, et al. 2010. Neuropathological heterogeneity in frontotemporal lobar degeneration with TDP-43 proteinopathy: a quantitative study of 94 cases using principal components analysis. *J. Neural Transm.* 117:227–39
- Bigio EH. 2008. Update on recent molecular and genetic advances in frontotemporal lobar degeneration. J. Neuropathol. Exp. Neurol. 67:635–48
- Dickson DW, Rademakers R, Hutton ML. 2007. Progressive supranuclear palsy: pathology and genetics. Brain Pathol. 17:74–82
- Joachim CL, Morris JH, Selkoe DJ. 1988. Clinically diagnosed Alzheimer's disease: autopsy results in 150 cases. Ann. Neurol. 24:50–56
- Brun A, Englund E. 1986. A white matter disorder in dementia of the Alzheimer type—a pathoanatomical study. Ann. Neurol. 19:253–62
- Nelson PT, Alafuzoff I, Bigio EH, Bouras C, Braak H, et al. 2012. Correlation of Alzheimer disease neuropathologic changes with cognitive status: a review of the literature. *J. Neuropathol. Exp. Neurol.* 71:362–81
- Braak H, Braak E. 1991. Neuropathological staging of Alzheimer related changes. Acta Neuropathol. 82:239–59
- 26. Braak H, Duyckaerts C, Braak E, Piette F. 1993. Neuropathological staging of Alzheimer-related changes with psychometrically assessed intellectual status. In *Alzbeimer's Disease: Advances in Clinical and Basic Research*, ed. B Corain, K Iqbal, M Nicolini, B Winblad, H Wisniewski, P Zatta, pp. 131–37. Chichester, UK: Wiley
- Braak H, Thal DR, Ghebremedhin E, Del Tredici K. 2011. Stages of the pathologic process in Alzheimer disease: age categories from 1 to 100 years. *J. Neuropathol. Exp. Neurol.* 70:960–69
- Masters CL, Simms G, Weinman NA, Multhaup G, McDonald BL, Beyreuther K. 1985. Amyloid plaque core protein in Alzheimer disease and Down syndrome. *PNAS* 82:4245–49
- 29. Dickson DW. 1997. The pathogenesis of senile plaques. J. Neuropathol. Exp. Neurol. 56:321-39
- Blessed G, Tomlinson BE, Roth M. 1968. The association between quantitative measures of dementia and of senile change in the cerebral grey matter of elderly subjects. Br. J. Psychiatry 117:797–811

- Berlau DJ, Kahle-Wrobleski K, Head E, Goodus M, Kim R, Kawas C. 2007. Dissociation of neuropathologic findings and cognition. Case report of an apolipoprotein E ε2/ε2 genotype. Arch. Neurol. 64:1193–96
- Serrano-Pozo A, Mielke ML, Muzitansky A, Gomez-Isla T, Growdon JH, et al. 2012. Stable size distribution of amyloid plaques over the course of Alzheimer disease. J. Neuropathol. Exp. Neurol. 71:694–701
- Thal DR, Rub U, Orantes M, Braak H. 2002. Phases of Aβ-deposition in the human brain and its relevance for the development of AD. *Neurology* 58:1791–800
- Vinters HV, Secor DL, Read SL, Frazee JG, Tomiyasu U, et al. 1994. Microvasculature in brain biopsy specimens from patients with Alzheimer's disease: an immunohistochemical and ultrastructural study. *Ultrastruct. Pathol.* 18:333–48
- 35. Kidd M. 1963. Paired helical filaments in electron microscopy of Alzheimer's disease. Nature 197:192-93
- Braak H, Braak E, Grundke-Iqbal I, Iqbal K. 1986. Occurrence of neuropil threads in the senile human brain and in Alzheimer's disease: a third location of paired helical filaments outside of neurofibrillary tangles and neuritic plaques. *Neurosci. Lett.* 65:351–55
- Braak H, Braak E. 1988. Neuropil threads occur in the dendrites of tangle-bearing nerve cells. Neuropathol. Appl. Neurobiol. 14:39–44
- Wisniewski K, Jervis GA, Moretz RC, Wisniewski HM. 1979. Alzheimer neurofibrillary tangles in disease other than senile and presenile dementia. *Ann. Neurol.* 5:288–94
- Mischel PS, Nguyen LP, Vinters HV. 1995. Cerebral cortical dysplasia associated with pediatric epilepsy. Review of neuropathologic features and proposal for a grading system. *J. Neuropathol. Exp. Neurol.* 54:137–53
- Duong T, DeRosa MJ, Poukens V, Vinters HV, Fisher RS. 1994. Neuronal cytoskeletal abnormalities in human cerebral cortical dysplasia. *Acta Neuropathol.* 87:493–503
- Tai HC, Serrano-Pozo A, Hashimoto T, Frosch MP, Spires-Jones TL, Hyman BT. 2012. The synaptic accumulation of hyperphosphorylated tau oligomers in Alzheimer disease is associated with dysfunction of the ubiquitin-proteasome system. *Am. J. Pathol.* 181:1426–35
- 42. Vinters HV. 1987. Cerebral amyloid angiopathy. A critical review. Stroke 18:311-24
- Vinters HV, Gilbert JJ. 1983. Cerebral amyloid angiopathy: incidence and complications in the aging brain. II. The distribution of amyloid vascular changes. Stroke 14:924–28
- Vinters HV, Wang ZZ, Secor DL. 1996. Brain parenchymal and microvascular amyloid in Alzheimer's disease. *Brain Pathol.* 6:179–95
- Ellis RJ, Olichney JM, Thal LJ, Mirra SS, Morris JC, et al. 1996. Cerebral amyloid angiopathy in the brains of patients with Alzheimer's disease. The CERAD experience, part XV. *Neurology* 46:1592–96
- Glenner GG, Wong CW. 1984. Alzheimer's disease: initial report of the purification and characterization of a novel cerebrovascular amyloid protein. *Biochem. Biophys. Res. Commun.* 120:885–890
- 47. Verbeek MM, de Waal RM, Vinters HV, eds. 2000. Cerebral Amyloid Angiopathy in Alzheimer's Disease and Related Disorders. Dordrecht, Neth.: Kluwer Acad.
- 48. Wang Z, Natte R, Berliner JA, Van Duinen SG, Vinters HV. 2000. Toxicity of Dutch (E22Q) and Flemish (A21G) mutant amyloid β proteins to human cerebral microvessel and aortic smooth muscle cells. Stroke 31:534–38
- Wang ZZ, Wu DF, Vinters HV. 2002. Hypoxia and reoxygenation of brain microvascular smooth muscle cells in vitro: cellular responses and expression of cerebral amyloid angiopathy–associated proteins. *Acta Pathol. Microbiol. Immunol. Scand.* 110:423–34
- Soontornniyomkij V, Choi C, Pomakian J, Vinters HV. 2010. High-definition characterization of cerebral β-amyloid angiopathy in Alzheimer's disease. *Hum. Pathol.* 41:1601–8
- Miyakawa T, Katsuragi S, Watanabe K, Shimoji A, Ikeuchi Y. 1986. Ultrastructural studies of amyloid fibrils and senile plaques in human brain. *Acta Neuropathol.* 70:202–8
- 52. Attems J, Yamaguchi H, Saido TC, Thal DR. 2010. Capillary CAA and perivascular Aβ-deposition: two distinct features of Alzheimer's disease pathology. *J. Neurol. Sci.* 299:155–62
- 53. Zabel M, Schrag M, Crofton A, Tung S, Beaufond P, et al. 2013. A shift in microglial β-amyloid binding in Alzheimer's disease is associated with cerebral amyloid angiopathy. *Brain Pathol.* 23:390–401

- 54. Weller RO, Yow HY, Preston SD, Mazanti I, Nicoll JAR. 2002. Cerebrovascular disease is a major factor in the failure of elimination of Aβ from the aging human brain: implications for therapy of Alzheimer's disease. Ann. N.Y. Acad. Sci. 977:162–68
- Herzig MC, Van Nostrand WE, Jucker M. 2006. Mechanism of cerebral β-amyloid angiopathy: murine and cellular models. *Brain Pathol.* 16:40–54
- Zhang-Nunes SX, Maat-Schieman MLC, Van Duinen SG, Roos RAC, Frosch MP, Greenberg SM. 2006. The cerebral β-amyloid angiopathies: hereditary and sporadic. *Brain Pathol.* 16:30–39
- 57. Haglund M, Passant U, Sjobeck E, Ghebremedhin E, Englund E. 2006. Cerebral amyloid angiopathy and cortical microinfarcts as putative substrates of vascular dementia. *Int. J. Geriatr. Psychiatry* 21:681–87
- Soontornniyomkij V, Lynch MD, Mermash S, Pomakian J, Badkoobehi H, et al. 2010. Cerebral microinfarcts associated with severe cerebral β-amyloid angiopathy. *Brain Pathol.* 20:459–67
- Anders KH, Wang ZZ, Kornfeld M, Gray V, Soontornniyomkij V, et al. 1997. Giant cell arteritis in association with cerebral amyloid angiopathy: immunohistochemical and molecular studies. *Hum. Pathol.* 28:1237–46
- Arvanitakis Z, Leurgans SE, Wang Z, Wilson RS, Bennett DA, Schneider JA. 2011. Cerebral amyloid angiopathy pathology and cognitive domains in older persons. *Ann. Neurol.* 69:320–27
- Ball MJ. 1977. Neuronal loss, neurofibrillary tangles and granulovacuolar degeneration in the hippocampus with ageing and dementia: a quantitative study. Acta Neuropathol. 37:111–18
- Ball MJ. 1978. Topographic distribution of neurofibrillary tangles and granulovacuolar degeneration in hippocampal cortex of aging and demented patients: a quantitative study. Acta Neuropathol. 42:73–80
- Clare R, King VG, Wirenfeldt M, Vinters HV. 2010. Synapse loss in dementias. J. Neurosci. Res. 88:2083– 90
- 64. Terry RD, Masliah E, Salmon DP, Butters N, DeTeresa R, et al. 1991. Physical basis of cognitive alterations in Alzheimer's disease: Synapse loss is the major correlate of cognitive impairment. Ann. Neurol. 30:572–80
- Davidsson P, Blennow K. 1998. Neurochemical dissection of synaptic pathology in Alzheimer's disease. Int. Psychogeriatr. 10:11–23
- Cleary JP, Walsh DM, Hofmeister JJ, Shankar GM, Kuskowski MA, et al. 2005. Natural oligomers of the amyloid-β protein specifically disrupt cognitive function. *Nat. Neurosci.* 8:79–84
- Selkoe DJ. 2008. Soluble oligomers of the amyloid β-protein impair synaptic plasticity and behavior. Behav. Brain Res. 192:106–13
- Walsh DM, Klyubin I, Fadeeva JV, Cullen WK, Anwyl R, et al. 2002. Naturally secreted oligomers of amyloid beta protein potently inhibit hippocampal long-term potentiation in vivo. *Nature* 416:535–39
- 69. Ihara M, Kalaria RN. 2007. Amyloid-β and synaptic activity in mice and men. NeuroReport 18:1205-6
- Matsuyama S, Teraoka R, Mori H, Tomiyama T. 2007. Inverse correlation between amyloid precursor protein and synaptic plasticity in transgenic mice. *NeuroReport* 18:1083–87
- Fu C, Chute DJ, Farag ES, Garakian J, Cummings JL, Vinters HV. 2004. Comorbidity in dementia: an autopsy study. Arch. Pathol. Lab. Med. 128:32–38
- 72. Gearing M, Mirra SS, Hedreen JC, Sumi SM, Hansen LA, Heyman A. 1995. The Consortium to Establish a Registry for Alzheimer's Disease (CERAD). Part X. Neuropathology confirmation of the clinical diagnosis of Alzheimer's disease. *Neurology* 45:461–66
- 73. Selnes OA, Vinters HV. 2006. Vascular cognitive impairment. Nat. Clin. Pract. Neurol. 2:538-47
- 74. Vinters HV, Ellis WG, Zarow C, Zaias BW, Jagust WJ, et al. 2000. Neuropathologic substrates of ischemic vascular dementia. *J. Neuropathol. Exp. Neurol.* 59:931–45
- Woodhouse A, Dickson TC, Vickers JC. 2007. Vaccination strategies for Alzheimer's disease: a new hope? Drugs Aging 24:107–19
- Karran E, Hardy J. 2014. Antiamyloid therapy for Alzheimer's disease—are we on the right road? N. Engl. J. Med. 370:377–78
- Vinters HV, Pardridge WM, Yang J. 1988. Immunohistochemical study of cerebral amyloid angiopathy: use of an antiserum to a synthetic 28-amino-acid peptide fragment of the Alzheimer's disease amyloid precursor. *Hum. Patbol.* 19:214–22

- Vinters HV, Nishimura GS, Secor DL, Pardridge WM. 1990. Immunoreactive A4 and gamma-trace peptide co-localization in amyloidotic arteriolar lesions in the brains of patients with Alzheimer's disease. *Am. 7. Pathol.* 137:233–40
- Galasko D, Hansen LA, Katzman R, Wiederholt W, Masliah E, et al. 1994. Clinical-neuropathological correlations in Alzheimer's disease and related dementias. *Arch. Neurol.* 51:888–95
- Di Patre PL, Read SL, Cummings JL, Tomiyasu U, Vartavarian LM, et al. 1999. Progression of clinical deterioration and pathological changes in patients with Alzheimer disease evaluated at biopsy and autopsy. *Arch. Neurol.* 56:1254–61
- 81. Khachaturian ZS. 1985. Diagnosis of Alzheimer's disease. Arch. Neurol. 42:1097-105
- Newell KL, Hyman BT, Growdon JH, Hedley-Whyte ET. 1999. Application of the National Institute on Aging (NIA)–Reagan Institute criteria for the neuropathological diagnosis of Alzheimer disease. *J. Neuropathol. Exp. Neurol.* 58:1147–55
- Postupna N, Rose SE, Bird TD, Gonzalez-Cuyar LF, Sonnen JA, et al. 2012. Novel antibody capture assay for paraffin-embedded tissue detects wide-ranging amyloid beta and paired helical filament-tau accumulation in cognitively normal older adults. *Brain Pathol.* 22:472–84
- Hyman BT, Phelps CH, Beach TG, Bigio EH, Cairns NJ, et al. 2012. National Institute on Aging– Alzheimer's Association guidelines for the neuropathologic assessment of Alzheimer's disease. *Alzheimer's Dement*. 8:1–13
- Montine TJ, Phelps CH, Beach TG, Bigio EH, Cairns NJ, et al. 2012. National Institute on Aging– Alzheimer's Association guidelines for the neuropathologic assessment of Alzheimer's disease: a practical approach. *Acta Neuropathol.* 123:1–11
- Ritchie K, Touchon J. 2000. Mild cognitive impairment: conceptual basis and current nosological status. Lancet 355:225–28
- Petersen RC, Parisi JE, Dickson DW, Johnson KA, Knopman DS, et al. 2006. Neuropathologic features of amnestic mild cognitive impairment. *Arch. Neurol.* 63:665–72
- 88. Vinters HV. 2006. Neuropathology of amnestic mild cognitive impairment. Arch. Neurol. 63:645-46
- Jicha GA, Parisi JE, Dickson DW, Johnson K, Cha R, et al. 2006. Neuropathologic outcome of mild cognitive impairment following progression to clinical dementia. *Arch. Neurol.* 63:674–81
- 90. Schneider JA, Arvanitakis Z, Leurgans SE, Bennett DA. 2009. The neuropathology of probable Alzheimer disease and mild cognitive impairment. *Ann. Neurol.* 66:200–8
- Shepherd C, McCann H, Halliday GM. 2009. Variations in the neuropathology of familial Alzheimer's disease. Acta Neuropathol. 118:37–52
- Jonsson T, Atwal JK, Steinberg S, Snaedel J, Jonsson PV, et al. 2012. A mutation in APP protects against Alzheimer's disease and age-related cognitive decline. Nature 488:96–99
- Arnold SE, Vega IE, Karlawish JH, Wolk DA, Nunez J, et al. 2013. Frequency and clinicopathological characterization of presenilin 1 Gly206Ala mutation in Puerto Rican Hispanics with dementia. *J. Alzbeimer's Dis.* 33:1089–95
- Holton P, Ryten M, Nalls M, Trabzuni D, Weale ME, et al. 2013. Initial assessment of the pathogenic mechanisms of the recently identified Alzheimer risk loci. *Ann. Hum. Genet.* 77:85–105
- Neumann H, Daly MJ. 2013. Variant TREM2 as risk factor for Alzheimer's disease. N. Engl. J. Med. 368:182–84
- 96. Reitz C, Mayeux R. 2013. TREM2 and neurodegenerative disease. N. Engl. J. Med. 369:1564-65
- 97. Bertram L, Parrado AR, Tanzi RE. 2013. Letter to the editor. N. Engl. J. Med. 369:1565
- 98. Jellinger KA, Attems J. 2007. Neuropathological evaluation of mixed dementia. J. Neurol. Sci. 257:80-87
- Robinson JL, Geser F, Corrada MM, Berlau DJ, Arnold SE, et al. 2011. Neocortical and hippocampal amyloid-β and tau measures associate with dementia in the oldest-old. *Brain* 134:3708–15
- 100. Sonnen JA, Larson EB, Haneuse S, Woltjer R, Crane PK, et al. 2009. Neuropathology in the Adult Changes in Thought study: a review. J. Alzheimer's Dis. 18:703–11
- 101. Schneider JA, Aggarwal NT, Barnes L, Boyle P, Bennett DA. 2009. The neuropathology of older persons with and without dementia from community versus clinic cohorts. *J. Alzbeimer's Dis.* 18:691–701
- 102. Schneider JA, Bennett DA. 2010. Where vascular meets neurodegenerative disease. *Stroke* 41(Suppl. 1):S144-46

- Hachinski V, Iadecola C, Petersen RC, Breteler MM, Nyenhuis D, et al. 2006. National Institute of Neurological Disorders and Stroke–Canadian Stroke Network vascular cognitive impairment harmonization standards. *Stroke* 37:2220–41
- 104. Jack CR Jr. 2011. Alliance for Aging Research AD Biomarkers Work Group: structural MRI. Neurobiol. Aging 32:S48–57
- 105. Jagust W. 2013. Biomarkers and brain connectivity. JAMA Neurol. 70:1233-34
- Schott JM, Fox NC, Frost C, Scahill RI, Janssen JC, et al. 2003. Assessing the onset of structural change in familial Alzheimer's disease. *Ann. Neurol.* 53:181–88
- 107. Chan D, Fox NC, Jenkins R, Scahill RI, Crum WR, et al. 2001. Rates of global and regional cerebral atrophy in AD and frontotemporal dementia. *Neurology* 57:1756–63
- Erten-Lyons D, Dodge HH, Woltjer R, Silbert LC, Howleson DB, et al. 2013. Neuropathologic basis of age-associated brain atrophy. *JAMA Neurol.* 70:616–22
- 109. Klunk WE, Engler H, Nordberg A, Wang Y, Blomqvist G, et al. 2004. Imaging brain amyloid in Alzheimer's disease with Pittsburgh Compound-B. Ann. Neurol. 55:306–19
- Weiner MW, Veitch DP, Aisen PS, Beckett LA, Cairns NJ, et al. 2012. The Alzheimer's disease Neuroimaging Initiative: a review of papers published since its inception. *Alzheimer's Dement*. 9:e111–94
- 111. Wirth M, Madison CM, Rabinovici GD, Oh H, Landau SM, Jagust WJ. 2013. Alzheimer's disease neurodegenerative biomarkers are associated with decreased cognitive function but not β-amyloid in cognitively normal older individuals. *J. Neurosci.* 33:5553–63
- 112. Wolk DA, Price JC, Madeira C, Saxton JA, Snitz BE, et al. 2012. Amyloid imaging in dementias with atypical presentation. *Alzbeimer's Dement.* 8:389–98
- Sanchez-Juan P, Ghosh PM, Hagen J, Gesierich B, Henry M, et al. 2014. Practical utility of amyloid and FDG-PET in an academic dementia center. *Neurology* 82:230–38
- 114. Reiman EM, Jagust WJ. 2012. Brain imaging in the study of Alzheimer's disease. NeuroImage 61:505-16
- Oh H, Madison C, Villeneuve S, Markley C, Jagust WJ. 2014. Association of gray matter atrophy with age, β-amyloid, and cognition in aging. *Cereb. Cortex* 24:1609–18
- 116. Ikonomovic MD, Abrahamson EE, Price JC, Hamilton RL, Mathis CA, et al. 2012. Early AD pathology in a [C-11]PiB-negative case: a PiB-amyloid imaging, biochemical and immunohistochemical study. Acta Neuropathol. 123:433–47
- 117. Ikonomovic MD, Klunk WE, Abrahamson EE, Mathis CA, Price JC, et al. 2008. Post-mortem correlates of in vivo PiB-PET amyloid imaging in a typical case of Alzheimer's disease. *Brain* 131:1630–45
- Clark CM, Pontecorvo MJ, Beach TG, Bedell BJ, Coleman RE, et al. 2012. Cerebral PET with florbetapir compared with neuropathology at autopsy for detection of neuritic amyloid-β plaques: a prospective cohort study. *Lancet Neurol.* 11:669–78
- Lo RY, Jagust WJ, Alzheimer's Dis. Neuroimaging Initiat. 2013. Effect of cognitive reserve markers on Alzheimer pathologic progression. *Alzheimer Dis. Assoc. Disord.* 27:343–50
- Irwin DJ, Trojanowski JQ, Grossman M. 2013. Cerebrospinal fluid biomarkers for differentiation of frontotemporal lobar degeneration from Alzheimer's disease. *Front. Aging Neurosci.* 5:6
- 121. Ghidoni R, Benussi L, Paterlini A, Albertini V, Binetti G, Emanuele E. 2011. Cerebrospinal fluid biomarkers for Alzheimer's disease: the present and future. *Neurodegener. Dis.* 8:413–20
- 122. Landau SM, Lu M, Joshi AD, Pontecorvo M, Mintun MA, et al. 2013. Comparing positron emission tomography imaging and cerebrospinal fluid measurements of β-amyloid. Ann. Neurol. 74:826–36
- 123. Graff-Radford NR, Crook JE, Lucas J, Boeve BF, Knopman DS, et al. 2007. Association of low plasma Aβ42/Aβ40 ratios with increased imminent risk for mild cognitive impairment and Alzheimer disease. *Arcb. Neurol.* 64:354–62
- 124. Ray S, Britschgi M, Herbert C, Takeda-Uchimura Y, Boxer A, et al. 2007. Classification and prediction of clinical Alzheimer's diagnosis based on plasma signaling proteins. *Nat. Med.* 13:1359–62
- 125. Sunderland T, Hampel H, Takeda M, Putnam KT, Cohen RM. 2006. Biomarkers in the diagnosis of Alzheimer's disease: Are we ready? *J. Geriatr. Psychiatry Neurol.* 19:172–79
- 126. Freeman SH, Raju S, Hyman BT, Frosch MP, Irizarry MC. 2007. Plasma Aβ levels do not reflect brain Aβ levels. J. Neuropathol. Exp. Neurol. 66:264–71
- 127. Nicoll JAR, Wilkinson D, Holmes C, Steart P, Markham H, Weller RO. 2003. Neuropathology of human Alzheimer disease after immunization with amyloid-β peptide: a case report. *Nat. Med.* 9:448–52

- 128. Ferrer I, Rovira MB, Guerra MLS, Rey MJ, Costa-Jussa F. 2004. Neuropathology and pathogenesis of encephalitis following amyloid-β immunization in Alzheimer's disease. *Brain Pathol.* 14:11–20
- 129. Holmes C, Boche D, Wilkinson D, Yadegarfar G, Hopkins V, et al. 2008. Long-term effects of Aβ₄₂ immunisation in Alzheimer's disease: follow-up of a randomised, placebo-controlled phase I trial. *Lancet* 372:216–23
- Nicoll JAR, Barton E, Boche D, Neal JW, Ferrer I, et al. 2006. Aβ species removal after Aβ42 immunization. *J. Neuropathol. Exp. Neurol.* 65:1040–48
- Boche D, Denham N, Holmes C, Nicoll JAR. 2010. Neuropathology after active Aβ42 immunotherapy: implications for Alzheimer's disease pathogenesis. *Acta Neuropathol.* 120:369–84
- 132. Boche D, Zotova E, Weller RO, Love S, Neal JW, et al. 2008. Consequence of Aβ immunization on the vasculature of human Alzheimer's disease brain. *Brain* 131:3299–310
- 133. Maarouf CL, Daugs ID, Kokjohn TA, Kalback WM, Patton RL, et al. 2010. The biochemical aftermath of anti-amyloid immunotherapy. *Mol. Neurodegener.* 5:39
- 134. Zotova E, Bharambe V, Cheaveau M, Morgan W, Holmes C, et al. 2013. Inflammatory components in human Alzheimer's disease and after active amyloid-β₄₂ immunization. *Brain* 136:2677–96
- 135. Doody RS, Thomas RG, Farlow M, Iwatsubo T, Vellas B, et al. 2014. Phase 3 trials of solanezumab for mild-to-moderate Alzheimer's disease. *N. Engl. J. Med.* 370:311–21
- Salloway S, Sperling R, Fox NC, Blennow K, Klunk W, et al. 2014. Two phase 3 trials of bapineuzumab in mild-to-moderate Alzheimer's disease. N. Engl. J. Med. 370:322–33
- 137. Klyubin I, Walsh DM, Lemere CA, Cullen WK, Shankar GM, et al. 2005. Amyloid β protein immunotherapy neutralizes Aβ oligomers that disrupt synaptic plasticity in vivo. *Nat. Med.* 11:556–61
- 138. Miners JS, Barua N, Kehoe PG, Gill S, Love S. 2011. Aβ-degrading enzymes: potential for treatment of Alzheimer disease. *J. Neuropathol. Exp. Neurol.* 70:944–59
- Hulette C, Nochlin D, McKeel D, Morris JC, Mirra SS, et al. 1997. Clinical-neuropathologic findings in multi-infarct dementia: a report of six autopsied cases. *Neurology* 48:668–72
- Jellinger KA. 2007. The enigma of vascular cognitive disorder and vascular dementia. Acta Neuropathol. 113:349–88
- 141. Jellinger KA. 2008. The pathology of "vascular dementia": a critical update. J. Alzheimer's Dis. 14:107-23
- 142. Knopman DS. 2007. Cerebrovascular disease and dementia. Br. J. Radiol. 80:S121-27
- Chui HC, Zarow C, Mack WJ, Ellis WG, Zheng L, et al. 2006. Cognitive impact of subcortical vascular and Alzheimer's disease pathology. *Ann. Neurol.* 60:677–87
- 144. Gorelick PB, Scuteri A, Black SE, DeCarli C, Greenberg SM, et al. 2011. Vascular contributions to cognitive impairment and dementia: a statement for healthcare professionals from the American Heart Association/American Stroke Association. Stroke 42:2672–713
- Iadecola C. 2010. The overlap between neurodegenerative and vascular factors in the pathogenesis of dementia. Acta Neuropathol. 120:287–96
- 146. Jellinger KA, Attems J. 2010. Prevalence of dementia disorders in the oldest-old: an autopsy study. Acta Neuropathol. 119:421–33
- Langa KM, Foster NL, Larson EB. 2004. Mixed dementia: emerging concepts and therapeutic implications. *JAMA* 292:2901–8
- 148. Kovari E, Herrmann FR, Hof PR, Bouras C. 2013. The relationship between cerebral amyloid angiopathy and cortical microinfarcts in brain aging and Alzheimer's disease. *Neuropathol. Appl. Neurobiol.* 39:498– 509
- Thal DR, Ghebremedhin E, Orantes M, Wiestler OD. 2003. Vascular pathology in Alzheimer disease: correlation of cerebral amyloid angiopathy and arteriosclerosis/lipohyalinosis with cognitive decline. *J. Neuropathol. Exp. Neurol.* 62:1287–301
- 150. Thal DR, Grinberg LT, Attems J. 2012. Vascular dementia: Different forms of vessel disorders contribute to the development of dementia in the elderly brain. *Exp. Gerontol.* 47:816–24
- 151. Arvanitakis Z, Leurgans SE, Barnes LL, Bennett DA, Schneider JA. 2011. Microinfarct pathology, dementia, and cognitive systems. *Stroke* 42:722–27
- 152. Longstreth WT, Sonnen JA, Koepsell TD, Kukull WA, Larson EB, Montine TJ. 2009. Associations between microinfarcts and other macroscopic vascular findings on neuropathologic examination in 2 databases. *Alzheimer's Dis. Assoc. Disord*. 23:291–94

- 153. Dickson DW, Davies P, Bevona C, Van Hoeven KH, Factor SM, et al. 1994. Hippocampal sclerosis: a common pathological feature of dementia in very old (≥80 years of age) humans. Acta Neuropathol. 88:212–21
- Rauramaa T, Pikkarainen M, Englund E, Ince PG, Jellinger K, et al. 2011. Cardiovascular diseases and hippocampal infarcts. *Hippocampus* 21:281–87
- 155. Zarow C, Sitzer TE, Chui HC. Understanding hippocampal sclerosis in the elderly: epidemiology, characterization, and diagnostic issues. Curr. Neurol. Neurosci. Rep. 8:363–70
- Amador-Ortiz C, Ahmed Z, Zehr C, Dickson DW. 2007. Hippocampal sclerosis dementia differs from hippocampal sclerosis in frontal lobe degeneration. *Acta Neuropathol.* 113:245–52
- 157. Zarow C, Vinters HV, Ellis WG, Weiner MW, Mungas D, et al. 2005. Correlates of hippocampal neuron number in Alzheimer's disease and ischemic vascular dementia. *Ann. Neurol.* 57:896–903
- Ball MJ, Fisman M, Hachinski V, Blume W, Fox A, et al. 1985. A new definition of Alzheimer's disease: a hippocampal dementia. *Lancet* 325:14–16
- Deane R, Zlokovic BV. 2007. Role of the blood-brain barrier in the pathogenesis of Alzheimer's disease. Curr. Alzbeimer Res. 4:191–97
- Zlokovic BV. 2005. Neurovascular mechanisms of Alzheimer's neurodegeneration. *Trends Neurosci.* 28:202–8
- Jeynes B, Provias J. 2011. The case for blood-brain barrier dysfunction in the pathogenesis of Alzheimer's disease. J. Neurosci. Res. 89:22–28
- Stewart PA, Hayakawa K, Akers MA, Vinters HV. 1992. A morphometric study of the blood-brain barrier in Alzheimer's disease. *Lab. Investig.* 67:734–42
- 163. Beach TG, Wilson JR, Sue LI, Newell A, Poston M, et al. 2007. Circle of Willis atherosclerosis: association with Alzheimer's disease, neuritic plaques and neurofibrillary tangles. Acta Neuropathol. 113:13–21
- 164. Honig LS, Kukull W, Mayeux R. 2005. Atherosclerosis and AD: analysis of data from the US National Alzheimer's Coordinating Center. *Neurology* 64:494–500
- 165. Luoto TM, Haikonen S, Haapasalo H, Goebeler S, Huhtala H, et al. 2009. Large vessel atherosclerosis is not in direct association with neuropathological lesions of Alzheimer's disease. *Eur. Neurol.* 62:93–98
- 166. Yarchoan M, Xie SX, Kling MA, Toledo JB, Wolk DA, et al. 2012. Cerebrovascular atherosclerosis correlates with Alzheimer pathology in neurodegenerative dementias. *Brain* 135:3749–56
- 167. Dolan H, Crain B, Troncoso J, Resnick SM, Zonderman AB, O'Brien RJ. 2010. Atherosclerosis, dementia, and Alzheimer disease in the Baltimore Longitudinal Study of Aging cohort. Ann. Neurol. 68:231–40
- 168. Chui HC, Zheng L, Reed BR, Vinters HV, Mack WJ. 2012. Vascular risk factors and Alzheimer's disease: Are these risk factors for plaques and tangles or for concomitant vascular pathology that increases the likelihood of dementia? An evidence-based review. *Alzheimer's Res. Ther.* 4:1
- Zheng L, Vinters HV, Mack WJ, Zarow C, Ellis WG, Chui HC. 2013. Cerebral atherosclerosis is associated with cystic infarcts and microinfarcts but not Alzheimer pathologic changes. *Stroke* 44:2835– 41
- Rockenstein E, Crews L, Masliah E. 2007. Transgenic animal models of neurodegenerative diseases and their application to treatment development. *Adv. Drug Deliv. Rev.* 59:1093–102
- Schellenberg GD, Montine TJ. 2012. The genetics and neuropathology of Alzheimer's disease. Acta Neuropathol. 124:305–23
- 172. Huang Y, Mucke L. 2012. Alzheimer mechanisms and therapeutic strategies. Cell 148:1204-22
- 173. Ball MJ, Lukiw WJ, Kammerman EM, Hill JM. 2013. Intracerebral propagation of Alzheimer's disease: strengthening evidence of a herpes simplex virus etiology. *Alzheimer's Dement*. 9:169–75
- 174. Aguzzi A, Rajendran L. 2009. The transcellular spread of cytosolic amyloids, prions, and prionoids. *Neuron* 64:783–90
- 175. Stohr J, Watts JC, Mensinger ZL, Oehler A, Grillo SK, et al. 2012. Purified and synthetic Alzheimer's amyloid beta (Aβ) prions. PNAS 109:11025–30
- 176. Hamaguchi T, Eisele YS, Varvel NH, Lamb BT, Walker LC, Jucker M. 2012. The presence of Aβ seeds, and not age *per se*, is critical to the initiation of Aβ deposition in the brain. *Acta Neuropathol.* 123:31–37
- 177. Harris JA, Devidze N, Verret L, Ho K, Halabisky B, et al. 2010. Transsynaptic progression of amyloidβ-induced neuronal dysfunction within the entorhinal-hippocampal network. *Neuron* 68:428–41

- Braak H, Del Tredici K. 2011. Alzheimer's pathogenesis: Is there neuron-to-neuron propagation? Acta Neuropathol. 121:589–95
- 179. Clavaguera F, Bolmont T, Crowther A, Abramowski D, Frank S, et al. 2009. Transmission and spread of tauopathy in transgenic mouse brain. *Nat. Cell Biol.* 11:909–14
- Castellani RJ, Perry G. 2014. The complexities of the pathology-pathogenesis relationship in Alzheimer disease. *Biochem. Pharmacol.* 88:671–76
- 181. Nunomura A, Perry G, Aliev G, Hirai K, Takeda A, et al. 2001. Oxidative damage is the earliest event in Alzheimer disease. *7. Neuropathol. Exp. Neurol.* 60:759–67
- Kovacs GG, Budka H. 2010. Current concepts of neuropathological diagnostics in practice: neurodegenerative diseases. *Clin. Neuropathol.* 29:271–88
- Kumar-Singh S, Van Broeckhoven C. 2007. Frontotemporal lobar degeneration: current concepts in the light of recent advances. *Brain Pathol.* 17:104–13
- McKeith IG, Dickson DW, Lowe J, Emre M, O'Brien JT, et al. 2005. Diagnosis and management of dementia with Lewy bodies: third report of the DLB consortium. *Neurology* 65:1863–72
- Wilson RS, Leurgans SE, Boyle PA, Schneider JA, Bennett DA. 2010. Neurodegenerative basis of agerelated cognitive decline. *Neurology* 75:1070–78
- van Swieten J, Spillantini MG. 2007. Hereditary frontotemporal dementia caused by *Tau* gene mutations. Brain Pathol. 17:63–73