

Primary Atopic Disorders

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Annu. Rev. Immunol. 2020. 38:785–808

First published as a Review in Advance on
March 3, 2020

The *Annual Review of Immunology* is online at
immunol.annualreviews.org

<https://doi.org/10.1146/annurev-immunol-042718-041553>

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Keywords

allergy, genetics, primary atopic disorders, monogenic, Th2, mast cell function, skin barrier

Abstract

Primary atopic disorders describes a series of monogenic diseases that have allergy- or atopic effector-related symptoms as a substantial feature. The underlying pathogenic genetic lesions help illustrate fundamental pathways in atopy, opening up diagnostic and therapeutic options for further study in those patients, but ultimately for common allergic diseases as well. Key pathways affected in these disorders include T cell receptor and B cell receptor signaling, cytokine signaling, skin barrier function, and mast cell function, as well as pathways that have not yet been elucidated. While comorbidities such as classically syndromic presentation or immune deficiency are often present, in some cases allergy alone is the presenting symptom, suggesting that commonly encountered allergic diseases exist on a spectrum of monogenic and complex genetic etiologies that are impacted by environmental risk factors.

INTRODUCTION

Much of the research on the genetics of atopic disease has focused on the hypothesis that allergy is a complex disease with common genetic underpinnings and specific environmental exposures. The pathways illuminated by this line of study have provided insight into the role of many of the genes anticipated by animal models of allergic effector biology (1). Another, nonexclusive, avenue that has begun to emerge more recently thanks to advances in genomic sequencing technology is the study of monogenic allergic diseases (2). These diseases can produce phenotypes much more striking in both severity and comorbidity than typical allergic disease, though the allergy can by and large be considered a severe form of typical allergic symptoms. The diseases also are remarkable for a lack of complete penetrance, providing potential opportunities to better study specific gene-environment interactions that could lead to allergic disease (3). While the number of patients with such disorders is not yet large enough to study on a population basis, it is an important open question whether the allergic phenotypes seen in monogenic diseases mirror common allergy in that the phenotypes have increased in recent generations as the result of Westernization.

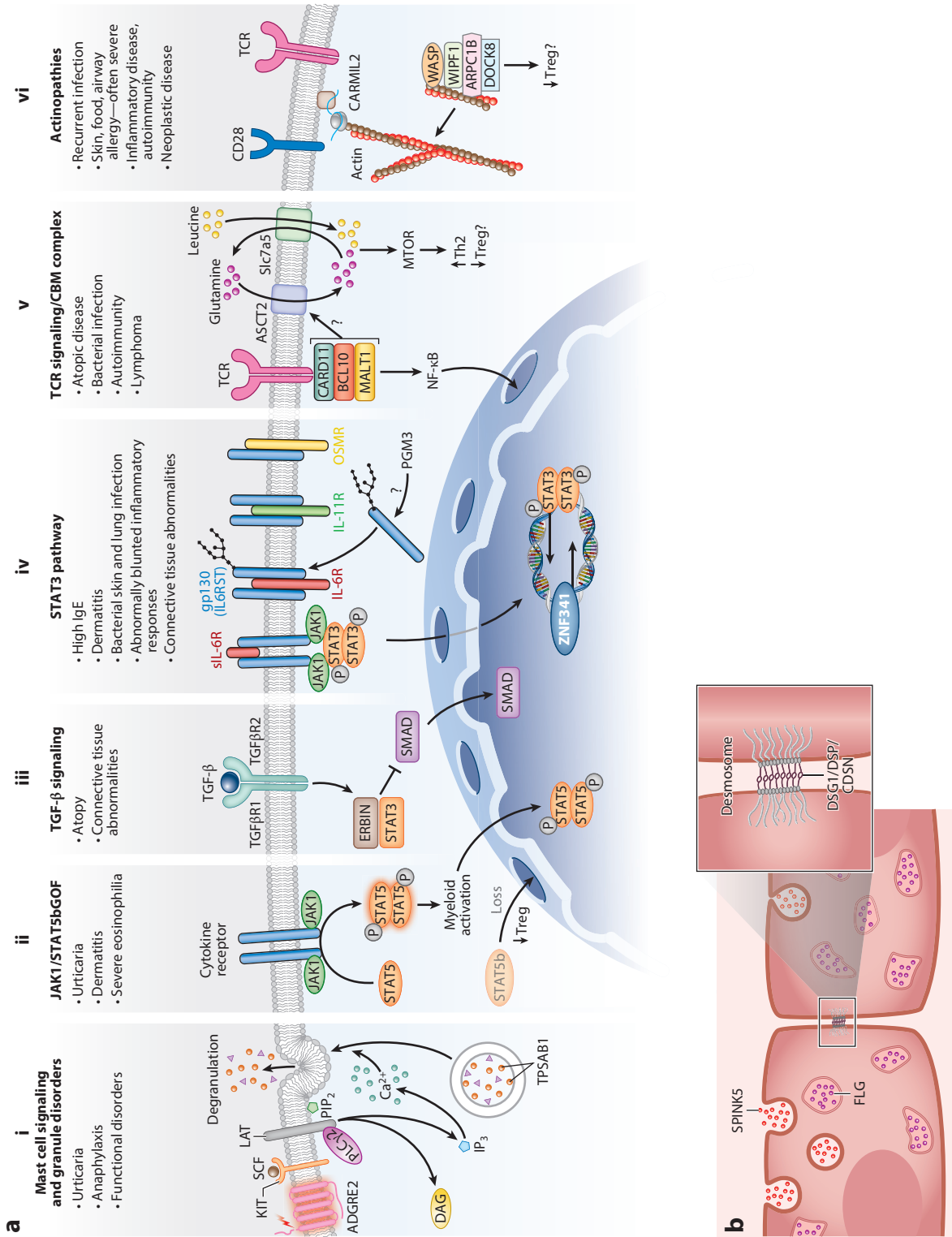
As these diseases have accumulated, a series of illuminating pathogenic pathways have coalesced—involving cytokine signaling, T cell receptor (TCR) signaling and actin dynamics, tolerance failure, intrinsic mast cell functions (**Figure 1a**), and physical barrier function (**Figure 1b**). In order to help distinguish allergic disease in the context of those with monogenic lesions from those that have other causes, the term primary atopic disorder (PAD) can be used to describe the condition of these patients (2). Patients with PADs may well have a primary immune deficiency as well, or a primary immune dysregulatory phenotype; however, to focus on allergic disease in these diseases of inborn errors of multiple pathways is critical both for patient care and understanding, as well as for research approaches.

In PAD cases where comorbid immune deficiency exists, the degree of impact of monogenic lesions on effector cell function can dictate the chances as to whether atopy will be present. As such, there are numerous examples of immune deficiencies so severe that hematopoietic stem cell transplantation is needed within the first year of life for survival, and in many cases, T cells may be lacking altogether, making it extremely difficult to mount an allergic response. However there are other examples of disease where mutations, often hypomorphic, allow for sufficient development of allergic effector cells but fail to fully prevent infection or pathogenic mechanisms that lead to allergic disease. Those disorders associated with substantial infection due to T cell defects, among others, are generally considered to be combined immune deficiencies (CID), to contrast with severe combined immune deficiencies (SCID), which result in the absence or near absence of T cell function. A subset of CID patients will develop atopic disease when the particular pathways that are partially ablated are normally responsible for preventing atopic disease (2).

In considering monogenic allergic disease, it is also important to separate the types of allergy one might observe, as they help define the clinical consequences of the underlying lesions and perhaps point to specific patterns of disease that can help identify the pathway or pathways involved. So for instance, marked IgE elevation might be associated with allergen-specific reactivity and defects of specific T and B cell tolerance pathways, chronic mucosal allergic inflammation such as eczema or eosinophilic gut disease might implicate barrier function defects, and urticaria or anaphylaxis in the absence of inflammation or antibody-mediated sensitization might point to aberrant intrinsic mast cell effector functions. The disorders described below are summarized in **Table 1**.

IMPAIRED TCR SIGNALING AND CYTOSKELETAL REMODELING

The association between allergic disease and immune deficiency is both across the board and focally enhanced in a number of disorders (4). One key determinant for whether allergic disease



(Caption appears on following page)

Figure 1 (Figure appears on preceding page)

Schematic representation of pathways involved in primary atopic disorders. (a, i) In mast cells, mechanical stress can more easily activate ADGRE2, leading to degranulation, while PLAID mutant PLC γ 2 can be activated by cold to lead to spontaneous calcium flux and degranulation. Increased alpha tryptase encoded by TPSAB1 leads to hereditary alpha tryptasemia. (ii) Activating mutations in JAK1 or STAT5b lead to myeloid-driven eosinophilia, dermatitis, and urticaria, while loss of STAT5b function leads to Treg defects and tolerance failure, which can include atopy. (iii) TGF β R1 or TGF β R2 activates SMADs, but ERBIN/STAT3 complexes normally prevent SMAD entry to the nucleus. Mutations of any of these genes are associated with connective tissue abnormalities and atopy. (iv) gp130 pairs with a variety of cytokine receptors to activate STAT3, which can upregulate ZNF341—which in turn upregulates STAT3. Loss of function along the entire pathway leads to distinct features of high IgE, atopy, and poor inflammatory responses. (v) The CBM complex is activated by antigen receptor and can control mTORC1 activation via control of glutamine transport. Hypomorphic mutations in this pathway can lead to viral skin infection, bacterial infection, and substantial atopic disease. (vi) Actin assembly downstream of TCR activation is critical for host defense and for adequate Treg function. Lesions in CARMIL2, WASP, WIPF1, DOCK8, and ARPC1B, all of which contribute to normal actin dynamics in one way or another, lead to recurrent infection in the pattern of combined immune deficiency, as well as substantial tolerance failures and atopic disease. (b) Structural proteins in keratinocytes such as filaggrin, or protease inhibitors such as SPINK5, as well as intercellular adhesion molecules such as CDSN, DSG1, or DSP all maintain barrier integrity in the outer layer of skin. Complete loss of function in any of them leads to ichthyosis or severe eczema, while milder mutations in FLG are risk factors for commonly encountered eczema. Abbreviations: CBM, CARD11/BCL10/MALT1; PLAID, PLC γ 2-associated antibody deficiency and immune dysregulation; TCR, T cell receptor; Treg, regulatory T cell; WASP, Wiskott-Aldrich syndrome protein.

Th2: type 2 T helper

Treg: regulatory
T cell

will be present in an immune deficiency is, as might be imagined, the presence of sufficient effector mechanisms to launch an atopic response. As such, patients lacking T cells who have SCID, for instance, will not be capable of developing allergic disease, lacking IgE or type 2 T helper (Th2) cells. Remaining allergic effectors such as eosinophils or type 2 innate lymphoid cells (ILC2s) do not appear to be sufficient to cause allergic pathology. However, when there is even a small leaky T cell population in patients with SCID, substantial homeostatic, oligoclonal expansion can occur, and the clinical presentation is termed Omenn syndrome (OS). These patients develop marked lymphoproliferation and organomegaly, elevated IgE despite the marked reduction in B cells, and severe inflammatory, erythrodermic skin disease with T cell infiltration that resembles the most severe cases of atopic dermatitis (5–8).

The allergic effector component of the OS phenotype has a number of potential explanations that may also apply to other disorders of limited TCR repertoire not as severe as OS. These include clonotypic gaps in regulatory T cell (Treg) specificity that cannot limit effector T cell responses (9–11); a lack of central tolerance due to poor upregulation of AIRE in the abnormal SCID thymus (12); and clonotypic gaps in high-affinity antigen-specific effector T cells, which

Table 1 Genetic mutations in primary atopic disorders

Altered process	Genes
Impaired TCR signaling and cytoskeletal remodeling	<i>CARD11, MALT1, WAS, WIPF1, ARPC1B, DOCK8, CARMIL2</i>
Altered cytokine signaling	<i>STAT3^{DN}, STAT1^{GOF}, STAT5B^{LOF}, STAT5B^{GOF}, JAK1^{GOF}, IL4RA^{GOF}, IL6ST, IL6R, TGFB1, TGFB2, ERBB2IP, ZNF341</i>
T cell repertoire restriction	<i>RAG1, RAG2, DCLRE1C, ADA, IL2RA, IL7RA, CHD7, LIG4, ZAP70, 22q11del</i>
Tolerance failure	<i>FOXP3, IL2RA, STAT5B^{LOF}, TGFB1, TGFB2, WAS, ARPC1B, MALT1, CARD11, STAT1^{GOF}</i>
Glycosylation	<i>PGM3</i>
Skin barrier disruption	<i>FLG, CDSN, DSG1, DSP, SPINK5</i>
Mast cell deregulation	<i>PLCG2, ADGRE2, TPSAB1</i>

Abbreviation: TCR, T cell receptor.

lead to weaker stimulation of a given antigen-specific naive T cell during priming, resulting in Th2 skewing (13), to be discussed further below.

In many ways, the repertoire disturbances of OS are a special case of syndromic atopy in the sense that the allergic disease is not allergen specific due to the marked repertoire restriction. However one potential mechanistic overlap may exist between OS and other PADs with intrinsic defects, specifically in the realm of TCR signaling.

The initial strength of TCR-peptide-MHC (pMHC) interaction can strongly contribute to the ultimate T helper fate of the naive T cell, in addition to, if not superseding, other costimulatory and cytokine cues more typically associated with T helper differentiation (14–20). Indeed hypomorphic germ line mutations of TCR signaling molecules, in contrast to knockout mouse models, in ζ chain of TCR-associated protein kinase 70 (*Zap70*); linker for activation of T cells (*Lat*); caspase recruitment domain family, member 11 (*Card11*); and others can lead to profound spontaneous atopic phenotypes (21–24).

CARD11 is a member of the CARD11-BCL10 (B cell chronic lymphocytic leukemia/lymphoma 10)-MALT1 (mucosa-associated lymphoid tissue lymphoma translocation gene 1) (CBM) complex, which links certain lymphocyte receptor engagement with downstream NF- κ B and mTOR signaling (25, 26). Complete biallelic loss of CARD11 or BCL10 signaling leads to a combined immunodeficiency necessitating hematopoietic stem cell transplantation, while MALT1 loss of function (LOF) results in CID with variable expressivity of severe atopic dermatitis and other elements of tolerance loss such as inflammatory gut disease (27–32). Similar to that seen in the mouse, hypomorphic, dominant negative mutations in *CARD11* can result in substantial atopic dermatitis, elevated serum IgE, and other allergic phenotypes. Not all patients develop allergy, and in some cases, carriers have only allergic disease, which they eventually grow out of. There is also variably observed bacterial respiratory infection, viral skin infection, and autoimmunity, showing the pleiotropy of these types of mutations (33, 34).

The pathogenesis of allergic disease in MALT1 or CARD11 mutant patients may be multifactorial. Studies of hypomorphic CARD11 mouse models have suggested a numerical defect in Tregs due to poor TCR activation (24, 35). Further evidence for CBM complex-dependent MALT1 paracaspase activity in Tregs might also suggest a mechanism for loss of tolerance (36–43).

Patients with hypomorphic CARD11 mutations, however, do not have differences in Treg number ex vivo or suppressive capacity in vitro, though a focal Th2 regulatory defect cannot be ruled out.

An effector cell-intrinsic mechanism may also be at play. CARD11LOF T cells fail to activate ASCT2 for normal glutamine uptake after activation. In turn, the low intracellular glutamine pools fail to adequately activate mTORC1, a key metabolic regulator of T helper differentiation (44). Loss of mTORC1 component Rheb leads to poor Th1 and enhanced Th2 differentiation (45), and a similar phenotype can be found in ASCT2 mutant mice. MALT1 and CARD11 mutant patients both have impaired mTORC1 activation in addition to their NF- κ B defect. Human naive T cells differentiated in the absence of glutamine have a profound shift away from Th1 cytokine expression even under Th1-skewing conditions. GATA3 expression is enhanced as well in low-glutamine conditions (46). Poor ex vivo expression of IFN- γ and in vitro Th1 differentiation were also clearly observed in patients with mutations in CARD11. The extent to which the patients' Th2 phenotype emerges because of a further intrinsic Th2 bias of differentiating T cells, or simply the lack of counterregulating Th1 cytokines, requires further study.

The CBM complex activates NF- κ B, and another open question surrounds the role of impaired NF- κ B signaling in human atopic disease. The numerous monogenic disorders affecting NF- κ B components are not associated with atopic disease despite adequate effector function, suggesting that lesions upstream lead to allergy independently of NF- κ B. That said, a number of mouse

CBM complex:
CARD11-BCL10-
MALT1 complex

mTORC1:
mammalian target of
rapamycin complex 1

Th1: type 1 T helper

models at least suggest that NF- κ B pathway mutants could lead to allergic disease especially in the context of viral infection (47), and patients with mutations in EDAR1 and 2, which primarily signal through NF- κ B, can develop significant atopic dermatitis and other allergic phenotypes (48, 49).

The case of EDAR mutations is instructive since the defect is thought to be largely restricted to keratinocytes. While skin barrier defects are covered in a subsequent section, it is relevant to note here that mutations in another keratinocyte-restricted gene, *CARD14*, can lead to atopy as well. *CARD14* is structurally and functionally analogous to *CARD11*, forming the keratinocyte CBM complex. Dominant negative mutations in *CARD14* have been found in multiple patients with severe atopic dermatitis and skin infections. *CARD14* gain-of-function (GOF) mutations are known to lead to skin inflammatory diseases that resemble psoriasis, enhanced NF- κ B expression, antimicrobial peptide (AMP) expression, and autoexpression of *CARD14* protein. In contrast, *CARD14*LOF keratinocytes fail to normally activate NF- κ B, or upregulate *CARD14* and AMP expression (50). The loss versus gain of AMP expression mimics a key difference between more typical, common psoriasis and atopic dermatitis whereby AMP expression is excessive in psoriasis and impaired in atopic dermatitis. It is thought that in common atopic dermatitis, impaired endogenous AMP production is secondary to local Th2 cytokines downregulating expression, explaining the superinfections seen in atopic dermatitis skin, in contrast to inflamed psoriatic skin (51). Whether the primary AMP production defect in *CARD14* could directly lead to atopic dermatitis—perhaps via introduction of Th2 driving bacterial products—remains to be determined.

Intriguingly, the dermatitis and other mucosal inflammatory and infectious phenotypes seen in some *MALT1*LOF patients is fully cured with bone marrow transplantation (30, 52, 53). As such, while *MALT1* is expressed in skin keratinocytes, and *CARD14* is thought to signal through *MALT1* to upregulate NF- κ B and AMPs, the hematopoietic compartment alone appears to account for the atopic dermatitis and elevated IgE seen in *MALT1*LOF. It is also possible that the conditioning regimen for transplantation and reconstitution processes has some effect on risk for atopic dermatitis even in the context of a genetic lesion (54).

Another pathway that can affect TCR signaling and leads to substantial atopy when mutated relates to actin polymerization and cytoskeletal rearrangement. After TCR stimulation, Wiskott-Aldrich syndrome protein (WASP) is released by its inhibitor and stabilizer WASP-interacting protein (WIP) so that it is free to associate with the actin-related protein (ARP) 2/3 complex, leading to actin polymerization and subsequent structural changes associated with migration and proliferation (55).

LOF in X-linked *WASP* leads to Wiskott-Aldrich syndrome (WAS), classically characterized by severe atopic dermatitis, thrombocytopenia, combined immunodeficiency, and neoplastic disease (56). In addition to eczematous dermatitis, these patients also have a substantial predisposition toward food allergen sensitization and clinical food allergy (57). Other mutations in *WASP* lead to X-linked thrombocytopenia (XLT) and neutropenia. XLT can lead to a more limited atopic phenotype as well (57). Biallelic LOF in related actin polymerization genes (*WIPF1*) encoding WIP (58) and actin-related protein 2/3 complex subunit 1B (*ARPC1B*) (33, 59, 60) lead to very similar phenotypes to WAS, including the substantial atopic diathesis.

Another stabilizing target of WIP is dedicator of cytokinesis 8 (DOCK8)—a guanine nucleotide exchange factor whose activity is essential to normal WASP function (61). It is therefore not surprising that LOF mutations in *DOCK8* lead to a combined immunodeficiency remarkable for severe viral skin infection—which can lead to neoplastic disease—in addition to bacterial respiratory infection. Allergic disease in *DOCK8* deficiency is quite prevalent and severe (62), perhaps more so than WAS since loss of *DOCK8* activity may be compensated in mast cells (57, 63, 64).

Treg failure appears to be a key mechanism for atopic disease in these actinopathies, as Treg function has been shown to be impaired in patient cells (57, 65–68) and deletion of *WASP* specifically in Tregs alone leads to substantial allergic sensitization and even more allergen reactivity than in germ line *WASP* mutants. The decreased severity in germ line mice is likely due to the importance of *WASP* in normal mast cell function (64).

DOCK8 deficiency may also lead to an intrinsic predisposition toward Th2 differentiation and away from Th1 (69), while *WAS* T cells appear to have an intrinsic bias away from Th1 cells as well (70, 71), again raising the question as to whether loss of IFN- γ alone can be a key mechanism for atopic diathesis. *WASP* mutations even appear to hinder certain Th2 phenotypes as well (72), providing another potential explanation for the severity of Treg-restricted *WASP* deletion in the mouse model.

Another scaffold protein potentially linked directly to actin dynamics is capping protein regulator and myosin 1 linker 2 (*CARMIL2*), encoding RGD, leucine-rich repeat, tropomodulin and proline-rich-containing protein (RLTPR). RLTPR appears to be critical for CD28 signaling in human T cells by linking CD28 signaling to CARD11 (73), and LOF mutations in *CARMIL2* lead to a combined immunodeficiency that can be associated with substantial comorbid atopy including severe atopic dermatitis, elevated IgE, allergic asthma, food allergy, and cold urticaria. Furthermore, the affected patients also had reduced numbers of Tregs (74–76). Further investigation will hopefully clarify the relative roles of impaired CARD11 activation, tolerance failure, or other intrinsically disordered actin dynamics in the pathogenesis of the atopic phenotypes in these patients.

ALTERED CYTOKINE SIGNALING

While TCR signaling appears to confer intrinsic defects in differentiation and in tolerance leading to atopic disease in the context of bacterial respiratory infection and viral skin infection, the actions of cytokines upon recent stimulation of naive T cells of course have a major impact upon differentiation as well, in addition to the many other ways in which cytokines may impact effector cells of all sorts.

A Number of PADs with Altered Cytokine Signaling Have Emerged

Altered cytokine signaling pathways appear to be a rather fundamental cause of PADs. One of the most obvious lesions in which allergic disease would be expected is loss of IL-12 or IFN- γ signaling; these key Th1-related cytokines are well known to counterregulate Th2 and atopic responses in mouse models and in vitro differentiation. In humans, such disorders are associated with increased Th2 cytokine production (77). However, while allergic disease can be present, it is not particularly enriched in those affected patients reported (78) either because of a lack of substantial Th1 counterregulation of Th2 effector functions in vivo in humans, or because of the tremendous burden of opportunistic infections that may inhibit Th2 effector function in vivo in ways independent of Th1-related cytokines.

STAT3 Pathway Disorders

One major pathway that appears to substantially contribute to monogenic allergic disease is the STAT3 signaling pathway. Mutations of STAT3, as well as in upstream and downstream members of the pathway, lead to overlapping clinical entities. The most immunologically characteristic elements include bacterial infection of the mucosa, poor inflammatory responses, dermatitis, and

marked IgE elevation. Chronic mucocutaneous candidiasis is variably associated, while viral or mycobacterial infection is not particularly characteristic.

Dominant negative mutations in *STAT3* (*STAT3^{DN}*) lead to a disorder initially described as Job syndrome, then autosomal dominant hyper-IgE syndrome (AD-HIES) (79, 80). Patients can develop recurrent bacterial sinopulmonary infections leading to abnormal lung cysts, chronic mucocutaneous candidiasis, a newborn rash, chronic eczematoid dermatitis, and recurrent staphylococcal skin abscesses deemed cold due to the lack of the cardinal signs of inflammation—calor (warmth), rubor (redness), dolor (pain), and tumor (swelling) (81–83). In addition patients have substantial connective tissue abnormalities involving head and facial dysmorphism, easily broken bones, hypermobile joints, and arterial tortuosity and aneurysm formation. The marked IgE elevation, however, is what gave the disorder one of its names.

While patients tend not to have primary viral illnesses, herpesvirus reactivation such as varicella zoster virus recurrence and Epstein–Barr virus viremia does occur. Lymphomas, both Hodgkin and non-Hodgkin, are also enriched (84–86). The impaired chronic viral and tumor surveillance may at least in part be due to decreased circulating memory T cell formation, which is intrinsic to *STAT3^{DN}* (87, 88).

In addition to high IgE, *STAT3^{DN}* patients have peripheral eosinophilia and tissue infiltration of eosinophils, in particular in the gastrointestinal tract (89). Nonetheless, despite the elevated IgE and eosinophilia, patients with *STAT3^{DN}* are relatively protected from IgE-mediated food allergy and anaphylaxis (87, 90). This protection mirrors the global defect in inflammation seen in these patients, suggesting a common mechanism. However, a number of reasons have been posited for the poor allergic responses, including failure to make adequate antigen-specific IgE, a role for *STAT3* in normal mast cell degranulation, and a role for *STAT3* in normal vascular endothelial responses to histamine (87, 91, 92). The elevation in IgE itself is also not well understood. Th2 cytokines are enriched among the reduced number of memory T cells, but interestingly mouse models strongly suggest that the predisposition to IgE production is B cell intrinsic (93, 94). Notably, there is a developmental defect in B cell maturation in a number of PADs, and transitional B cells in particular have an intrinsic predisposition to IgE class-switching (94, 95). Furthermore, impaired TLR9 signaling in B cells, which appears to be dependent on both DOCK8 and *STAT3*, may contribute to intrinsic IgE class-switch predilection as well (96).

The IL-10 receptor utilizes *STAT3* for signaling transduction and tends to protect against many types of inflammation, and its absence leads to severe early-onset colitis. However, such disease is not present in *STAT3^{DN}* despite poor IL-10R signaling, potentially reflecting other defects in inflammatory cells and mediator responses. Certain types of autoimmunity have been reported, including systemic lupus erythematosus (97–99), autoimmune vasculitis, dermatomyositis, and membranoproliferative glomerulonephritis. Whether these diseases are truly enriched in the *STAT3^{DN}* cohort remains to be determined.

The connective tissue defects in *STAT3^{DN}* are important since they overlap with a number of other monogenic disorders to be discussed later in this review, some of which lead to allergic disease as well. These include retained primary teeth, easily fractured bones, joint hypermobility, craniosynostosis, characteristic facies, and scoliosis (82, 100, 101). Vascular abnormalities are seen in *STAT3^{DN}* as well, such as arterial tortuosity, dilation, and aneurysm formation (102–107).

STAT3 activation leads to expression of a series of target genes, which leads to a complex array of biological responses. Part of the responses are autoregulatory. SOCS3 upregulation, for instance, leads to inhibition of *STAT3* and other JAK-*STAT* signaling pathways. Interestingly, study of patients with ZNF341 mutation has shown that a positive feed-forward loop exists as well, whereby cytokine-induced *STAT3* activation leads to upregulation of *STAT3* expression itself via

the transcription factor ZNF341. Recessive ZNF341 LOF mutations lead to a phenotype quite similar to *STAT3^{DN}* including marked serum IgE elevation, bacterial skin and respiratory tract infections, fungal infections, and atypical inflammatory responses such as cold abscesses, as well as connective tissue abnormalities that were perhaps not as significant as those seen in *STAT3^{DN}* (108, 109).

Upstream of STAT3, a key cytokine coreceptor critical for signaling for a variety of cytokines including IL-6, IL-11, IL-27, OSMR, LIF, and others is gp130, encoded by *IL6ST*. gp130 can also signal via other STATs, and as such despite the very small sample size of recessive *IL6ST* LOF demonstrates phenotypic overlap with but also distinction from *STAT3^{DN}*. Bacterial infection, connective tissue abnormalities and high IgE were all noted in GP130 deficiency, as was diarrhea, keratitis, and neurodevelopmental delay in one of the patients. Again while the small sample size precludes judgment regarding the severity or type of allergic disease, the phenotype largely appears similar to *STAT3^{DN}*. Notable laboratory findings include poor upregulation of the inflammatory marker, CRP, in the context of infection, and quite similar to the case of *STAT3^{DN}*, low frequencies of memory T and B cells, mucosal-associated invariant T (MAIT) cells, and Th1 and Th17 cells (110, 111).

Given the overlap between gp130 deficiency and *STAT3^{DN}*, it stands to reason that loss of gp130-associated cytokine receptors would also contribute to disease. One might have imagined that poor responses to STAT3-dependent, gp130-independent cytokine IL-21 or IL-10 could explain the high IgE for a number of reasons; however, it is not clear that would be the case in humans. IL-21 deficiency in mice can lead to elevated baseline IgE (112), though not in all models, and in fact IL-21 may have an obligatory role in certain Th2 responses (113). *IL6^{-/-}* mice can develop elevated IgE despite a Th2 cytokine production defect, and it is postulated that this elevation is indirect, due to a secondary IL-21 production defect (114). IL-21 signaling may suppress IgE class-switching (115), possibly via ID2 induction, and prevent anaphylaxis (116). However in humans IL-21 signaling, likely through STAT3, appears critical at least for in vitro IgE class-switching (117), and patients who lack IL-21 or IL-21R may have had some mild elevation in IgE but nothing approaching the levels seen in HIES, and without a rash or other atopic phenotypes (118, 119). Similarly, IL-10 is thought to be critical in establishing tolerance to allergens (120); however, IL-10R deficiency leads to early-onset inflammatory bowel disease, but as mentioned above not IgE elevation or atopy (121).

Two unrelated patients with LOF mutations in the *IL6R* gene, encoding the IL-6 receptor, have been identified. The IL-6 receptor is the gp130 binding partner for IL-6-mediated signaling. The patients had atopic dermatitis, elevated IgE bacterial sinopulmonary infection, and substantial skin and soft tissue infections—often due to staphylococcus. As with gp130/STAT3/ZNF341 LOF, they had abnormal inflammatory responses, including the absence of elevations in CRP (122).

The findings in *IL6R* LOF may well clarify which gp130/STAT3-dependent cytokine is critical for suppressing IgE/atopy during development. Furthermore, these patients provide context to data showing that a common coding variant in *IL6R* that leads to diminution of function can predispose to asthma (123), elevated IgE levels in asthma (124), and persistent atopic dermatitis (125). The same variant actually protects from other types of inflammatory disease such as rheumatoid arthritis and type 1 diabetes. This is especially of interest since IL-6 receptor blockade is utilized in a number of such inflammatory diseases but does not lead to allergic disease, nor does the presence of naturally arising IL-6 autoantibodies, which are associated with staphylococcal cellulitis and skin abscesses (126), arguing that there is an early-life window when IL-6-mediated signaling is critical to prevent allergy.

STAT5B^{GOF}:

gain-of-function
STAT5B mutations

JAK1^{GOF}:

gain-of-function *JAK1*
mutations

EGID: eosinophilic
gastrointestinal disease

JAK1/STAT5b Pathway

The role for STAT5 signaling in atopic disease is complex. Somatic heterozygous GOF mutations in STAT5b within the hematopoietic compartment can lead to massive neonatal eosinophilia, urticaria, diarrhea, and granulomatous disease (127). The patients' phenotype was predicted in part by a series of experiments in mice lacking PLCB3, which appears to be a negative regulator of STAT5b. The increased STAT5 signaling in PLCB3 leads to a myeloproliferative atopic phenotype (128, 129). The exact STAT5b mutation in *STAT5B^{GOF}* has previously been reported in association with leukemia and lymphomas (130–132), some of which are associated with eosinophilia (133). It is therefore interesting that the neonatal-onset somatic mutation does not lead to overt neoplasm over the years, but rather what looks to be an inborn error leading to a PAD (127).

Further evidence for the contribution of GOF in this pathway is found in a report of a family with a GOF heterozygous mutation in JAK1 (*JAK1^{GOF}*) that can lead to STAT5B phosphorylation (134). Unlike the STAT5bGOF patients, carriers had severe atopic dermatitis; however, quite similar to STAT5bGOF patients, they had marked peripheral and tissue eosinophilia and allergic asthma. Treatment with ruxolitinib, a JAK inhibitor, led to marked clinical improvement and resolution of eosinophilia, highlighting the power of precision medicine whereby the pathogenic lesion could be identified and directly treated to resolve allergic disease.

It is relevant here to note that mutations in *TYK2*, which bridges cytokine receptor signaling to STATs similarly to JAKs, were described in a single patient with markedly elevated IgE and mycobacterial infection, and as such were posited to be a cause of hyper-IgE (135). The initial patient had a defect in IL-6 signaling, which would be consistent with the observation that IL-6 receptor LOF leads to IgE elevation. However subsequently, numerous patients were identified with *TYK2* deficiency and none had atopy or high IgE, nor was there an IL-6 receptor signaling defect based on ex vivo and transfection experiments, suggesting that the IL-6 defect was not intrinsic to *TYK2* in the initially reported patient (136).

TGF- β Pathway Mutations

In addition to substantial mouse data, abnormal TGF- β signaling has been implicated in a series of human allergic diseases including severe asthma and eosinophilic gastrointestinal disease (EGID) (137, 138). How exactly the normal signaling prevents allergic disease is not entirely clear. Clear LOF in *TGFB1* appears to lead to tolerance failure, though not in the form of allergic disease (139), while *TGFB1* GOF mutations, which lead to a number of developmental defects and connective tissue abnormalities, are not associated with allergic disease (140). In addition, increased TGF- β signaling as measured by SMADs has been observed in association with abnormal connective tissue phenotypes in certain atopic patients (141, 142). Perhaps most strikingly, heterozygous mutations in transforming growth factor beta receptor 1 and 2 (*TGFB1* and *TGFB2*) found in Loeys-Dietz syndrome lead to vascular and connective tissue abnormalities that strongly overlap with those seen in STAT3 pathway disorders (143) and are associated with substantial IgE-mediated food allergy, EGID, allergic asthma, and atopic dermatitis. These patients and mouse models of the disease are noted to have enhanced SMAD2/3 phosphorylation in primary cells ex vivo, though contributing further to the lack of clarity in how TGF- β might lead to allergic disease, the receptor mutation itself is thought to be LOF (144). Abnormal Th2 cytokine production by Foxp3⁺ T cells points toward a potential mechanism (145). Allergic disease has not yet been carefully measured in other mutations associated with altered TGF- β signaling or availability such as those with mutations in FBN, TNXB, SMAD3, TGFB2, TGFB3, and SKI (146–148).

While it is still unresolved how TGF- β signaling leads to allergic disease in the disorders above, one possible link between STAT3 pathway mutations and the TGF- β pathway may be found in

a family with a heterozygous LOF mutation in *erbb2*-interacting protein (*ERBB2IP*), encoding ERBIN. The patients presented with significant EGID, allergen-specific reactivity, and connective tissue abnormalities (149). When STAT3 induces and then forms a complex with ERBIN, SMAD2/3 nuclear localization is limited. However in lymphocytes from ERBIN or STAT3LOF patients, TGF- β pathway activation was enhanced, leading to increased IL4R α expression, a TGF- β target in certain cell types (150). IL4R α activation by TGF- β is a rather important potential mechanism for atopic phenotypes, since it is of course well known to lead to Th2 differentiation and IgE production, a common GOF *IL4RA* variant increases the risk for a number of allergic diseases (151), and IL4R α may have a direct role in mediating pruritus in neurons (152).

Impaired function of Tregs—alluded to above in relation to actinopathies, CBM complex formation, and others—is a fundamental mechanism that can lead to allergic diseases. X-linked loss of forkhead box P3 (*Foxp3*) function in male mice (153) describes the scurfy phenotype and in humans the immunodysregulation polyendocrinopathy enteropathy X-linked (IPEX) syndrome (154–156). While patients are best known for early-onset, severe autoimmunity, which can be fatal, they also can develop severe atopic dermatitis, allergen reactivity, high IgE, and eosinophilia (157). Treg function is dependent on IL-2 receptor signaling via STAT5b, and as such LOF in IL-2 coreceptor CD25 (158), encoded by *IL2RA*, and the LOF mutations in *STAT5B* can lead to phenotypes, including atopy, similar to IPEX. STAT5bLOF patients also have susceptibility to viral infection—likely due to poor IL-2-mediated effector functions—and short stature, likely due to roles for STAT5b in growth factor signaling as well (159, 160).

Why there is selectivity for Th2 dysregulation and not other forms of inflammation and autoimmunity in some PADs where Tregs are implicated is not well understood. There is in vivo precedence for the direct and specific role for FOXP3 in Th2 disease, however. Mice with specific deletion in the conserved noncoding sequence 1 (CNS1)—which has been shown to be critical for peripheral Treg identity in controlling FOXP3 expression—develop spontaneous allergic inflammation of the mucosal surfaces (161). Tregs from patients with a specific mutation in the domain swap interface of FOXP3 have a selective inability to control their own Th2 cytokine production, leading to a highly Th2-skewed phenotype in the patient; the same is true in a mouse model (162).

SKIN BARRIER DISRUPTION

Another fundamental pathway required to prevent allergic disease is skin barrier function. Multiple disorders have been described that affect the physical skin barrier and lead to severe atopic disease. Filaggrin, encoded by *FLG*, serves as a physical barrier protein in a number of ways, including being processed into natural moisturizing factor (NMF) (163). Biallelic LOF mutations in *FLG* cause a severe form of eczematous dermatitis, ichthyosis vulgaris (164). These extreme cases show the spectrum of consequences from altered filaggrin expression. LOF heterozygous *FLG* variants are highly associated with common atopic dermatitis and other allergic disease (165, 166). Filaggrin expression is actually lowered by Th2 cytokines, illustrating a vicious cycle in allergic disease (167, 168).

Another key structure in skin barrier function is the desmosome, a junctional protein complex that helps seal the keratinocyte barrier and resists mechanical stress (169, 170). Mutations in multiple subunits of the desmosome lead to structural abnormalities that cause barrier defects, and to severe atopic dermatitis and/or ichthyosis vulgaris including corneodesmosin (*CDSN*), desmoplakin (*DSP*), and desmoglein-1 (*DSG1*) (171–173). Biallelic LOF mutations in *CDSN* result in peeling skin syndrome, type B, which is characterized by diffuse ichthyosis and erythroderma of the skin, severe itching, marked IgE elevation and other symptomatic allergic disease (171). Perhaps even more severe are dominant mutations in *DSP* (172) and recessive LOF mutations in

DSG1 (173), which result in severe dermatitis, multiple allergies, and marked metabolic wasting, or SAM syndrome.

Desmosome turnover is a highly ordered process involving protease activity for degradation and protease inhibitors to regulate its rate. Lymphoepithelial Kazal-type-related inhibitor (LEKTI), encoded by serine protease inhibitor Kazal-type 5 (*SPINK5*), serves as a key protease inhibitor in desmosomes (174), and autosomal recessive LOF mutations in *SPINK5* lead to Comel-Netherton syndrome, characterized by destabilized desmosomes, excessive desquamation, severe ichthyosis and erythroderma, trichorrhexis invaginatam (bamboo hair), elevated IgE, and atopy, as well as recurrent infections, all of which may be responsive to intravenous immune globulin therapy (175).

How does defective barrier function lead specifically to allergic phenotypes? Proposed reasons include inappropriate allergen penetration of mucosal immune cells that are normally shielded from external antigens/allergens, leading to antigen presentation and reactivity; growth and penetration of microbiota and their toxins, such as *Staphylococcus aureus*, which has an adjuvant effect on allergic inflammation; and the simple water loss that leads to a milieu more conducive to pruritus and more inflamed, with scratching (176–178). Emollient therapy alone, in restoring barrier function without immunomodulation per se, might even prevent atopic dermatitis in infants (179).

MAST CELL DEREGULATION

Mast cell effector functions underlie many of the most commonly encountered allergic phenotypes. As such, monogenic disorders affecting mast cell function comprise a distinct subset of PADs, one of which tends to differ from many other types that are more commonly associated with comorbid immune deficiency or chronic inflammation such as atopic dermatitis, or even IgE elevation.

One such group is physical urticarias. Physical urticarias are quite striking, in that stimuli such as heat, cold, vibration, and even certain light wavelengths can trigger mast cell degranulation, most easily observed clinically in the form of wheals and flares. The overwhelming majority of patients with physical urticarias do not have them from birth, nor are they lifelong. Their cause remains largely elusive, though in the case of cold urticaria there is some evidence that autoimmunity could have a role (180). However in some cases a PAD has been identified that leads to a physical urticaria. One such example is PLC γ 2-associated antibody deficiency and immune dysregulation (PLAID) (181–183). Patients with PLAID develop pruritus and erythema with evaporative cooling—which is distinct from the typical cold urticaria, which is triggered by contact with cold objects such as ice. PLAID patients' urticarial symptoms are present from birth and are lifelong. Deletions of the autoinhibitory cSH2 domain underlie known cases of PLAID. These deletions appear to lead to cellular anergy at physiologic temperatures in cells that express substantial PLC γ 2—which is nearly all hematopoietic cells, with the notable exception of T cells. This anergic phenotype likely explains the poor B cell class-switching observed and comorbid humoral immune deficiency seen in a subset of PLAID patients (181–183). The slightest of subphysiologic temperatures leads to marked spontaneous PLC γ 2 activity (184)—leading to calcium flux and ERK activation—which in mast cells leads to degranulation without the presence of a ligand. Such activity occurs in other myeloid cells, which may explain the skin granulomas that can form in a subset of patients, likely due to the slight decrease in temperature at the body surfaces (183). The mechanism by which PLC γ 2 deletions lead to the complex signaling abnormalities likely affects its role as a scaffold protein in addition to the lack of autoinhibitory activity (185). The cold urticaria in PLAID is entirely distinct from the inflammatory rash that was confused for cold urticaria seen in familial cold-induced autoinflammatory disorders caused by

NLR family pyrin domain containing 3 (*NLRP3*) inflammasome mutations. The inflammasome rashes did appear urticarial, but they were neutrophilic, not mast cell related (186).

Vibratory stimuli can also physically trigger mast cell degranulation and urticaria. In one case, a founder population from Lebanon carried a mutation in *ADGRE2*, encoding EGF-like module-containing mucin-like hormone receptor-like 2 (EMR2), leading to lifelong vibratory urticaria with systemic symptoms when significant (187). EMR2 is a G protein-coupled receptor that binds to glycosaminoglycans such as dermatan sulfate. When physical shearing forces are applied—such as in vibration—a noncovalently bound subunit of the receptor dissociates, leading to cellular activation and mast cell degranulation, even in normal individuals, for reasons that are not well understood (188). The particular *ADGRE2* mutation in the Lebanese-descent cohort lowers the threshold for this physical dissociation and subsequent mast cell degranulation (187).

Finally, mast cell mediators themselves have genetically determined pathogenic roles leading to PADs. One such example is the case of familial alpha tryptasemia syndrome (189, 190). Tryptases are well known serine proteases found in mast cell granules. Their activation of PAR2 and other receptors has been posited to lead to a number of atopic and nonatopic phenotypes such as pruritus and tissue remodeling (191, 192), and elevations of basal serum tryptase are associated with a wide variety of symptoms, some typically associated with acute (such as flushing, itching, and anaphylaxis) and chronic (such as dysautonomia and fatigue) mast cell degranulation (193–197). There are two major secreted tryptase isoforms: alpha and beta tryptase. Secreted tryptases found in granules are typically packaged with heparin in a mature form as tetramers. Interestingly, alpha tryptase is catalytically inactive yet present in over two-thirds of the general population. Alpha/beta heterotetramers can form naturally, and have distinct properties and targets of action, including EMR2. Alpha/beta heterotetramers substantially increase the capacity for vibratory stimuli to dissociate EMR2, and patients with increased copy number of the alpha isoform of tryptase at *TPSAB1* are substantially more likely to experience urticaria with vibratory stimuli (198). Four to six percent of Caucasians carry these copy number increases, accounting for the vast majority of elevations in basal serum tryptase. These patients actually have increased basal levels of tryptase, most of which is alpha tryptase secreted in monomer form. Carriers of the increased alpha tryptase copy number have hereditary alpha tryptasemia (HAT), and those who are symptomatic with the above-described phenotypes have hereditary alpha tryptasemia syndrome (HATS). Whether these monomers also have a pathogenic role in some of the chronic symptoms seen in HATS remains to be seen.

ADDITIONAL PADs

A number of additional PADs not clearly linked to a specific pathogenic mechanism nonetheless provide opportunities for study of allergy pathogenesis. Autosomal recessive hypomorphic mutations in phosphoglucomutase 3 (*PGM3*) can lead to a multisystem disorder of infection, immune dysregulation, connective tissue abnormalities, and neurodevelopmental deficits. One element of the immune dysregulation is profound allergic disease including severe atopic dermatitis, food allergy, immediate and delayed hypersensitivity to medications, EGID, asthma, seasonal allergy, allergic bronchopulmonary aspergillosis, allergic fungal mastoiditis, and food-protein-induced enteropathy syndrome (199–203). *PGM3* is a critical enzyme in the hexosamine pathway required for normal production of uridine diphosphate *N*-acetylglucosamine (UDP-GlcNAc) (202, 204), which is essential for normal glycosylation. Complete absence of *PGM3* is not consistent with life, and as such these are hypomorphic mutations that lead to poor cytosolic UDP-GlcNAc production, which in turn leads to measurably lower glycosylation in certain tissues, in particular naive T cells (202, 204). How this leads to allergic disease is not understood. IgE glycosylation

itself appears to be normal (205). One potential mechanism suggests that adequate *N*-glycosylation of gp130 is required for normal surface expression, resulting in impaired signaling by gp130-dependent cytokines, which could explain some overlap of PGM3 deficiency with *IL6ST* LOF patients. The finding is complicated by the fact that PGM3-deficient patients have mostly effector memory T cells in the periphery, which normally have low gp130 expression. However it is an example of what may be a multifactorial pathogenesis, raising the question as to which proteins must be normally glycosylated to prevent atopic disease.

Another disease that is related to marked IgE elevation and some other atopic features is prolydase deficiency. Autosomal recessive LOF mutations in *PEPD* (206) lead to dermatitis and skin ulcerations, recurrent respiratory infections and mucosal inflammation, hepatosplenomegaly, elevated IgE, facial dysmorphism, and developmental delay (207–209). Prolidase is a dipeptidase involved in collagen breakdown whose absence leads to the accumulation of dipeptides (210). While poor type I interferon signaling, which can theoretically oppose atopy, has been noted (211), other patients with type I interferon defects have not been noted to have allergic disease.

CONCLUSION

The increasing number of PADs now recognized have either affirmed or revealed the relevance of multiple biological pathways in the pathogenesis of allergy. In some cases more than one pathway may be affected, which could well reflect that multiple pathways that lead to (or should prevent) atopic phenotypes could stem from nodal genetic programs. It is critical to note that the PADs described are ones that have been identified. There are likely many more monogenic allergic disorders that have not been identified that likely involve both known and novel pathogenic mechanisms. Furthermore, as the case of *CARD11* deficiency and others can illustrate, commonly appearing allergic disease may well be monogenic in some cases. The lesions and pathways identified can impact allergic disease in the general population in a variety of ways, providing evidence that common allergic disease can also benefit from precision medicine, which incorporates genetic findings and identifies affected pathways to design rational individualized therapies.

DISCLOSURE STATEMENT

The author is not aware of any affiliations, memberships, funding, or financial holdings that might be perceived as affecting the objectivity of this review.

ACKNOWLEDGMENTS

This work was supported by the Division of Intramural Research of the National Institute of Allergy and Infectious Diseases, NIH.

LITERATURE CITED

1. Bonnelykke K, Sparks R, Waage J, Milner JD. 2015. Genetics of allergy and allergic sensitization: common variants, rare mutations. *Curr. Opin. Immunol.* 36:115–26
2. Lyons JJ, Milner JD. 2018. Primary atopic disorders. *J. Exp. Med.* 215:1009–22
3. Sacco KA, Milner JD. 2019. Gene-environment interactions in primary atopic disorders. *Curr. Opin. Immunol.* 60:148–55
4. Sokol K, Milner JD. 2018. The overlap between allergy and immunodeficiency. *Curr. Opin. Pediatr.* 30:848–54
5. Villa A, Notarangelo LD, Roifman CM. 2008. Omenn syndrome: inflammation in leaky severe combined immunodeficiency. *J. Allergy Clin. Immunol.* 122:1082–86

6. Joshi AY, Ham EK, Shah NB, Dong X, Khan SP, Abraham RS. 2012. Atypical Omenn syndrome due to adenosine deaminase deficiency. *Case Rep. Immunol.* 2012:919241
7. Turul T, Tezcan I, Artac H, de Bruin-Versteeg S, Barendregt BH, et al. 2009. Clinical heterogeneity can hamper the diagnosis of patients with ZAP70 deficiency. *Eur. J. Pediatr.* 168:87–93
8. Markert ML, Alexieff MJ, Li J, Sarzotti M, Ozaki DA, et al. 2004. Complete DiGeorge syndrome: development of rash, lymphadenopathy, and oligoclonal T cells in 5 cases. *J. Allergy Clin. Immunol.* 113:734–41
9. Milner JD, Ward JM, Keane-Myers A, Paul WE. 2007. Lymphopenic mice reconstituted with limited repertoire T cells develop severe, multiorgan, Th2-associated inflammatory disease. *PNAS* 104:576–81
10. Haribhai D, Williams JB, Jia S, Nickerson D, Schmitt EG, et al. 2011. A requisite role for induced regulatory T cells in tolerance based on expanding antigen receptor diversity. *Immunity* 35:109–22
11. Lawrence MG, Barber JS, Sokolic RA, Garabedian EK, Desai AN, et al. 2013. Elevated IgE and atopy in patients treated for early-onset ADA-SCID. *J. Allergy Clin. Immunol.* 132:1444–46
12. Cavadini P, Vermi W, Facchetti F, Fontana S, Nagafuchi S, et al. 2005. AIRE deficiency in thymus of 2 patients with Omenn syndrome. *J. Clin. Invest.* 115:728–32
13. Milner JD, Fazilleau N, McHeyzer-Williams M, Paul W. 2010. Cutting edge: lack of high affinity competition for peptide in polyclonal CD4⁺ responses unmasks IL-4 production. *J. Immunol.* 184:6569–73
14. Constant SL, Bottomly K. 1997. Induction of Th1 and Th2 CD4⁺ T cell responses: the alternative approaches. *Annu. Rev. Immunol.* 15:297–322
15. Yamane H, Paul WE. 2013. Early signaling events that underlie fate decisions of naive CD4⁺ T cells toward distinct T-helper cell subsets. *Immunol. Rev.* 252:12–23
16. Hosken NA, Shibuya K, Heath AW, Murphy KM, O'Garra A. 1995. The effect of antigen dose on CD4⁺ T helper cell phenotype development in a T cell receptor- $\alpha\beta$ -transgenic model. *J. Exp. Med.* 182:1579–84
17. Tao X, Grant C, Constant S, Bottomly K. 1997. Induction of IL-4-producing CD4⁺ T cells by antigenic peptides altered for TCR binding. *J. Immunol.* 158:4237–44
18. Jorritsma PJ, Brogdon JL, Bottomly K. 2003. Role of TCR-induced extracellular signal-regulated kinase activation in the regulation of early IL-4 expression in naive CD4⁺ T cells. *J. Immunol.* 170:2427–34
19. Barber JS, Yokomizo LK, Sheikh V, Freeman AF, Garabedian E, et al. 2013. Peptide library-based evaluation of T-cell receptor breadth detects defects in global and regulatory activation in human immunologic diseases. *PNAS* 110:8164–69
20. van Panhuys N, Klauschen F, Germain RN. 2014. T-cell-receptor-dependent signal intensity dominantly controls CD4⁺ T cell polarization in vivo. *Immunity* 41:63–74
21. Jakob T, Kollisch GV, Howaldt M, Bewersdorff M, Rathkolb B, et al. 2008. Novel mouse mutants with primary cellular immunodeficiencies generated by genome-wide mutagenesis. *J. Allergy Clin. Immunol.* 121:179–84.e7
22. Mingueneau M, Roncagalli R, Gregoire C, Kissenpfennig A, Miazek A, et al. 2009. Loss of the LAT adaptor converts antigen-responsive T cells into pathogenic effectors that function independently of the T cell receptor. *Immunity* 31:197–208
23. Jun JE, Wilson LE, Vinuesa CG, Lesage S, Blery M, et al. 2003. Identifying the MAGUK protein Carma-1 as a central regulator of humoral immune responses and atopy by genome-wide mouse mutagenesis. *Immunity* 18:751–62
24. Policheni A, Horikawa K, Milla L, Kofler J, Bouillet P, et al. 2019. CARD11 is dispensable for homeostatic responses and suppressive activity of peripherally induced FOXP3⁺ regulatory T cells. *Immunol. Cell Biol.* 97:740–52
25. Bertin J, Wang L, Guo Y, Jacobson MD, Poyet JL, et al. 2001. CARD11 and CARD14 are novel caspase recruitment domain (CARD)/membrane-associated guanylate kinase (MAGUK) family members that interact with BCL10 and activate NF- κ B. *J. Biol. Chem.* 276:11877–82
26. Hamilton KS, Phong B, Corey C, Cheng J, Gorentla B, et al. 2014. T cell receptor-dependent activation of mTOR signaling in T cells is mediated by Carma1 and MALT1, but not Bcl10. *Sci. Signal.* 7:ra55

27. Greil J, Rausch T, Giese T, Bandapalli OR, Daniel V, et al. 2013. Whole-exome sequencing links caspase recruitment domain 11 (CARD11) inactivation to severe combined immunodeficiency. *J. Allergy Clin. Immunol.* 131:1376–83.e3
28. Stepensky P, Keller B, Buchta M, Kienzler AK, Elpeleg O, et al. 2013. Deficiency of caspase recruitment domain family, member 11 (CARD11), causes profound combined immunodeficiency in human subjects. *J. Allergy Clin. Immunol.* 131:477–85.e1
29. Jabara HH, Ohsumi T, Chou J, Massaad MJ, Benson H, et al. 2013. A homozygous mucosa-associated lymphoid tissue 1 (MALT1) mutation in a family with combined immunodeficiency. *J. Allergy Clin. Immunol.* 132:151–58
30. Punwani D, Wang H, Chan AY, Cowan MJ, Mallott J, et al. 2015. Combined immunodeficiency due to MALT1 mutations, treated by hematopoietic cell transplantation. *J. Clin. Immunol.* 35:135–46
31. Torres JM, Martinez-Barricarte R, Garcia-Gomez S, Mazariegos MS, Itan Y, et al. 2014. Inherited BCL10 deficiency impairs hematopoietic and nonhematopoietic immunity. *J. Clin. Investig.* 124:5239–48
32. Frizinsky S, Rechavi E, Barel O, Najeeb RH, Greenberger S, et al. 2019. Novel *MALT1* mutation linked to immunodeficiency, immune dysregulation, and an abnormal T cell receptor repertoire. *J. Clin. Immunol.* 39:401–13
33. Somech R, Lev A, Lee YN, Simon AJ, Barel O, et al. 2017. Disruption of thrombocyte and T lymphocyte development by a mutation in *ARPC1B*. *J. Immunol.* 199:4036–45
34. Dadi H, Jones TA, Merico D, Sharfe N, Ovadia A, et al. 2017. Combined immunodeficiency and atopy caused by a dominant negative mutation in caspase activation and recruitment domain family member 11 (CARD11). *J. Allergy Clin. Immunol.* 141:1818–30.e2
35. Altin JA, Tian L, Liston A, Bertram EM, Goodnow CC, Cook MC. 2011. Decreased T-cell receptor signaling through CARD11 differentially compromises forkhead box protein 3-positive regulatory versus T_H2 effector cells to cause allergy. *J. Allergy Clin. Immunol.* 127:1277–85.e5
36. Barnes MJ, Krebs P, Harris N, Eidenschenk C, Gonzalez-Quintal R, et al. 2009. Commitment to the regulatory T cell lineage requires CARMA1 in the thymus but not in the periphery. *PLOS Biol.* 7:e51
37. Bornancin F, Renner F, Touil R, Sic H, Kolb Y, et al. 2015. Deficiency of MALT1 paracaspase activity results in unbalanced regulatory and effector T and B cell responses leading to multiorgan inflammation. *J. Immunol.* 194:3723–34
38. Brustle A, Brenner D, Knobbe-Thomsen CB, Cox M, Lang PA, et al. 2017. MALT1 is an intrinsic regulator of regulatory T cells. *Cell Death Differ.* 24:1214–23
39. Di Pilato M, Kim EY, Cadilha BL, Prussmann JN, Nasrallah MN, et al. 2019. Targeting the CBM complex causes Treg cells to prime tumours for immune checkpoint therapy. *Nature* 570:112–16
40. Medoff BD, Sandall BP, Landry A, Nagahama K, Mizoguchi A, et al. 2009. Differential requirement for CARMA1 in agonist-selected T-cell development. *Eur. J. Immunol.* 39:78–84
41. Molinero LL, Yang J, Gajewski T, Abraham C, Farrar MA, Alegre ML. 2009. CARMA1 controls an early checkpoint in the thymic development of FoxP3⁺ regulatory T cells. *J. Immunol.* 182:6736–43
42. Schmidt-Supprian M, Tian J, Grant EP, Pasparakis M, Maehr R, et al. 2004. Differential dependence of CD4⁺CD25⁺ regulatory and natural killer-like T cells on signals leading to NF- κ B activation. *PNAS* 101:4566–71
43. Rosenbaum M, Gewies A, Pechloff K, Heuser C, Engleitner T, et al. 2019. Bcl10-controlled Malt1 paracaspase activity is key for the immune suppressive function of regulatory T cells. *Nat. Commun.* 10:2352
44. Nakaya M, Xiao Y, Zhou X, Chang JH, Chang M, et al. 2014. Inflammatory T cell responses rely on amino acid transporter ASCT2 facilitation of glutamine uptake and mTORC1 kinase activation. *Immunity* 40:692–705
45. Delgoffe GM, Pollizzi KN, Waickman AT, Heikamp E, Meyers DJ, et al. 2011. The kinase mTOR regulates the differentiation of helper T cells through the selective activation of signaling by mTORC1 and mTORC2. *Nat. Immunol.* 12:295–303
46. Klysz D, Tai X, Robert PA, Craveiro M, Cretenet G, et al. 2015. Glutamine-dependent α -ketoglutarate production regulates the balance between T helper 1 cell and regulatory T cell generation. *Sci. Signal.* 8:ra97

47. Liu Y, Wang Z, De La Torre R, Barling A, Tsujikawa T, et al. 2017. Trim32 deficiency enhances Th2 immunity and predisposes to features of atopic dermatitis. *J. Investig. Dermatol.* 137:359–66
48. Mark BJ, Becker BA, Halloran DR, Bree AF, Sindwani R, et al. 2012. Prevalence of atopic disorders and immunodeficiency in patients with ectodermal dysplasia syndromes. *Ann. Allergy Asthma Immunol.* 108:435–38
49. Tuano KS, Orange JS, Sullivan K, Cunningham-Rundles C, Bonilla FA, Davis CM. 2015. Food allergy in patients with primary immunodeficiency diseases: prevalence within the US Immunodeficiency Network (USIDNET). *J. Allergy Clin. Immunol.* 135:273–75
50. Peled A, Sarig O, Sun G, Samuelov L, Ma CA, et al. 2019. Loss-of-function mutations in caspase recruitment domain-containing protein 14 (*CARD14*) are associated with a severe variant of atopic dermatitis. *J. Allergy Clin. Immunol.* 143:173–81.e10
51. Nomura I, Goleva E, Howell MD, Hamid QA, Ong PY, et al. 2003. Cytokine milieu of atopic dermatitis, as compared to psoriasis, skin prevents induction of innate immune response genes. *J. Immunol.* 171:3262–69
52. Charbit-Henrion F, Jeverica AK, Begue B, Markelj G, Parlato M, et al. 2017. Deficiency in mucosa-associated lymphoid tissue lymphoma translocation 1: a novel cause of IPEX-like syndrome. *J. Pediatr. Gastroenterol. Nutr.* 64:378–84
53. Rozmus J, McDonald R, Fung SY, Del Bel KL, Roden J, et al. 2016. Successful clinical treatment and functional immunological normalization of human MALT1 deficiency following hematopoietic stem cell transplantation. *Clin. Immunol.* 168:1–5
54. McKinnon ML, Rozmus J, Fung SY, Hirschfeld AF, Del Bel KL, et al. 2014. Combined immunodeficiency associated with homozygous MALT1 mutations. *J. Allergy Clin. Immunol.* 133:1458–62.e7
55. Zigmund SH. 2000. How WASP regulates actin polymerization. *J. Cell Biol.* 150:F117–20
56. Ochs HD. 2009. Mutations of the Wiskott-Aldrich Syndrome Protein affect protein expression and dictate the clinical phenotypes. *Immunol. Res.* 44:84–88
57. Lexmond WS, Goettel JA, Lyons JJ, Jacobse J, Deken MM, et al. 2016. FOXP3⁺ Tregs require WASP to restrain Th2-mediated food allergy. *J. Clin. Investig.* 126:4030–44
58. Lanzi G, Moratto D, Vairo D, Masneri S, Delmonte O, et al. 2012. A novel primary human immunodeficiency due to deficiency in the WASP-interacting protein WIP. *J. Exp. Med.* 209:29–34
59. Kuijpers TW, Tool ATJ, van der Bijl I, de Boer M, van Houdt M, et al. 2017. Combined immunodeficiency with severe inflammation and allergy caused by ARPC1B deficiency. *J. Allergy Clin. Immunol.* 140:273–77.e10
60. Kahr WH, Pluthero FG, Elkadri A, Warner N, Drobac M, et al. 2017. Loss of the Arp2/3 complex component ARPC1B causes platelet abnormalities and predisposes to inflammatory disease. *Nat. Commun.* 8:14816
61. Janssen E, Tohme M, Hedayat M, Leick M, Kumari S, et al. 2016. A DOCK8-WIP-WASp complex links T cell receptors to the actin cytoskeleton. *J. Clin. Investig.* 126:3837–51
62. Zhang Q, Davis JC, Lamborn IT, Freeman AF, Jing H, et al. 2009. Combined immunodeficiency associated with DOCK8 mutations. *N. Engl. J. Med.* 361:2046–55
63. Ogawa K, Tanaka Y, Uruno T, Duan X, Harada Y, et al. 2014. DOCK5 functions as a key signaling adaptor that links FcεRI signals to microtubule dynamics during mast cell degranulation. *J. Exp. Med.* 211:1407–19
64. Pivniouk VI, Snapper SB, Kettner A, Alenius H, Laouini D, et al. 2003. Impaired signaling via the high-affinity IgE receptor in Wiskott-Aldrich syndrome protein-deficient mast cells. *Int. Immunol.* 15:1431–40
65. Alroqi FJ, Charbonnier LM, Keles S, Ghandour F, Mouawad P, et al. 2017. DOCK8 deficiency presenting as an IPEX-like disorder. *J. Clin. Immunol.* 37:811–19
66. Humblet-Baron S, Sather B, Anover S, Becker-Herman S, Kasproicz DJ, et al. 2007. Wiskott-Aldrich syndrome protein is required for regulatory T cell homeostasis. *J. Clin. Investig.* 117:407–18
67. Maillard MH, Cotta-de-Almeida V, Takeshima F, Nguyen DD, Michetti P, et al. 2007. The Wiskott-Aldrich syndrome protein is required for the function of CD4⁺CD25⁺Foxp3⁺ regulatory T cells. *J. Exp. Med.* 204:381–91

68. Marangoni F, Trifari S, Scaramuzza S, Panaroni C, Martino S, et al. 2007. WASP regulates suppressor activity of human and murine CD4⁺CD25⁺FOXP3⁺ natural regulatory T cells. *J. Exp. Med.* 204:369–80
69. Tangye SG, Pillay B, Randall KL, Avery DT, Phan TG, et al. 2017. Deducator of cytokinesis 8-deficient CD4⁺ T cells are biased to a T_H2 effector fate at the expense of T_H1 and T_H17 cells. *J. Allergy Clin. Immunol.* 139:933–49
70. Taylor MD, Sadhukhan S, Kottangada P, Ramgopal A, Sarkar K, et al. 2010. Nuclear role of WASp in the pathogenesis of dysregulated T_H1 immunity in human Wiskott-Aldrich syndrome. *Sci. Transl. Med.* 2:37ra44
71. Trifari S, Sitia G, Aiuti A, Scaramuzza S, Marangoni F, et al. 2006. Defective Th1 cytokine gene transcription in CD4⁺ and CD8⁺ T cells from Wiskott-Aldrich syndrome patients. *J. Immunol.* 177:7451–61
72. Morales-Tirado V, Sojka DK, Katzman SD, Lazarski CA, Finkelman FD, et al. 2010. Critical requirement for the Wiskott-Aldrich syndrome protein in Th2 effector function. *Blood* 115:3498–507
73. Roncagalli R, Cucchetti M, Jarmuzynski N, Gregoire C, Bergot E, et al. 2016. The scaffolding function of the RLTPR protein explains its essential role for CD28 co-stimulation in mouse and human T cells. *J. Exp. Med.* 213:2437–57
74. Wang Y, Ma CS, Ling Y, Bousfiha A, Camcioglu Y, et al. 2016. Dual T cell- and B cell-intrinsic deficiency in humans with biallelic *RLTPR* mutations. *J. Exp. Med.* 213:2413–35
75. Alazami AM, Al-Helale M, Alhissi S, Al-Saud B, Alajlan H, et al. 2018. Novel CARMIL2 mutations in patients with variable clinical dermatitis, infections, and combined immunodeficiency. *Front. Immunol.* 9:203
76. Schober T, Magg T, Laschinger M, Rohlf M, Linhares ND, et al. 2017. A human immunodeficiency syndrome caused by mutations in CARMIL2. *Nat. Commun.* 8:14209
77. Tangye SG, Pelham SJ, Deenick EK, Ma CS. 2017. Cytokine-mediated regulation of human lymphocyte development and function: insights from primary immunodeficiencies. *J. Immunol.* 199:1949–58
78. Wood PM, Fieschi C, Picard C, Ottenhoff TH, Casanova JL, Kumararatne DS. 2005. Inherited defects in the interferon-gamma receptor or interleukin-12 signalling pathways are not sufficient to cause allergic disease in children. *Eur. J. Pediatr.* 164:741–47
79. Holland SM, DeLeo FR, Elloumi HZ, Hsu AP, Uzel G, et al. 2007. STAT3 mutations in the hyper-IgE syndrome. *N. Engl. J. Med.* 357:1608–19
80. Minegishi Y, Saito M, Tsuchiya S, Tsuge I, Takada H, et al. 2007. Dominant-negative mutations in the DNA-binding domain of STAT3 cause hyper-IgE syndrome. *Nature* 448:1058–62
81. Davis SD, Schaller J, Wedgwood RJ. 1966. Job's syndrome: recurrent, “cold”, staphylococcal abscesses. *Lancet* 287:1013–15
82. Buckley RH, Wray BB, Belmaker EZ. 1972. Extreme hyperimmunoglobulinemia E and undue susceptibility to infection. *Pediatrics* 49:59–70
83. Buckley RH. 2001. The hyper-IgE syndrome. *Clin. Rev. Allergy Immunol.* 20:139–54
84. Gorin LJ, Jeha SC, Sullivan MP, Rosenblatt HM, Shearer WT. 1989. Burkitt's lymphoma developing in a 7-year-old boy with hyper-IgE syndrome. *J. Allergy Clin. Immunol.* 83:5–10
85. Kashef MA, Kashef S, Handjani F, Karimi M. 2006. Hodgkin lymphoma developing in a 4.5-year-old girl with hyper-IgE syndrome. *Pediatr. Hematol. Oncol.* 23:59–63
86. Chandesaris MO, Melki I, Natividad A, Puel A, Fieschi C, et al. 2012. Autosomal dominant STAT3 deficiency and hyper-IgE syndrome: molecular, cellular, and clinical features from a French national survey. *Medicine* 91:e1–19
87. Siegel AM, Stone KD, Cruse G, Lawrence MG, Olivera A, et al. 2013. Diminished allergic disease in patients with STAT3 mutations reveals a role for STAT3 signaling in mast cell degranulation. *J. Allergy Clin. Immunol.* 132:1388–96
88. Ives ML, Ma CS, Palendira U, Chan A, Bustamante J, et al. 2013. Signal transducer and activator of transcription 3 (STAT3) mutations underlying autosomal dominant hyper-IgE syndrome impair human CD8⁺ T-cell memory formation and function. *J. Allergy Clin. Immunol.* 132:400–11.e9
89. Arora M, Bagi P, Strongin A, Heimall J, Zhao X, et al. 2017. Gastrointestinal manifestations of STAT3-deficient hyper-IgE syndrome. *J. Clin. Immunol.* 37:695–700

90. Boos AC, Hagl B, Schlesinger A, Halm BE, Ballenberger N, et al. 2014. Atopic dermatitis, STAT3- and DOCK8-hyper-IgE syndromes differ in IgE-based sensitization pattern. *Allergy* 69:943–53
91. Hox V, O'Connell MP, Lyons JJ, Sackstein P, Dimaggio T, et al. 2016. Diminution of signal transducer and activator of transcription 3 signaling inhibits vascular permeability and anaphylaxis. *J. Allergy Clin. Immunol.* 138:187–99
92. Erlich TH, Yagil Z, Kay G, Peretz A, Migalovich-Sheikhet H, et al. 2014. Mitochondrial STAT3 plays a major role in IgE-antigen-mediated mast cell exocytosis. *J. Allergy Clin. Immunol.* 134:460–69
93. Dascani P, Ding C, Kong X, Tieri D, Hu X, et al. 2018. Transcription factor STAT3 serves as a negative regulator controlling IgE class switching in mice. *Immunoborizons* 2:349–62
94. Kane A, Lau A, Brink R, Tangye SG, Deenick EK. 2016. B-cell-specific STAT3 deficiency: insight into the molecular basis of autosomal-dominant hyper-IgE syndrome. *J. Allergy Clin. Immunol.* 138:1455–58.e3
95. Wesemann DR, Magee JM, Boboila C, Calado DP, Gallagher MP, et al. 2011. Immature B cells preferentially switch to IgE with increased direct S μ to S ϵ recombination. *J. Exp. Med.* 208:2733–46
96. Massaad MJ, Cangemi B, Al-Herz W, LeFranc G, Freeman A, et al. 2017. DOCK8 and STAT3 dependent inhibition of IgE isotype switching by TLR9 ligation in human B cells. *Clin. Immunol.* 183:263–65
97. Schopfer K, Feldges A, Baerlocher K, Parisot RF, Wilhelm JA, Matter L. 1983. Systemic lupus erythematosus in *Staphylococcus aureus* hyperimmunoglobulinaemia E syndrome. *Br. Med. J.* 287:524–26
98. Jacobs DH, Macher AM, Handler R, Bennett JE, Collen MJ, Gallin JI. 1984. Esophageal cryptococcosis in a patient with the hyperimmunoglobulin E-recurrent infection (Job's) syndrome. *Gastroenterology* 87:201–3
99. Leyh F, Wendt V, Scherer R. 1986. [Systemic lupus erythematosus and hyper-IgE syndrome in a 13-year-old child]. *Z. Hautkr.* 61:611–14 (In German)
100. Borges WG, Hensley T, Carey JC, Petrak BA, Hill HR. 1998. The face of Job. *J. Pediatr.* 133:303–5
101. Grimbacher B, Holland SM, Gallin JI, Greenberg F, Hill SC, et al. 1999. Hyper-IgE syndrome with recurrent infections—an autosomal dominant multisystem disorder. *N. Engl. J. Med.* 340:692–702
102. Falah O, Thwaites SE, Chalmers RT. 2012. Ruptured thoracoabdominal aneurysm in a 27-year-old with hyper IgE syndrome. *J. Vasc. Surg.* 55:830–32
103. Kim Y, Nard JA, Saad A, Casselman J, Wessell KR, et al. 2015. Cerebral aneurysm in a 12-year-old boy with a STAT3 mutation (hyper-IgE syndrome). *Ann. Allergy Asthma Immunol.* 114:430–31
104. Ling JC, Freeman AF, Gharib AM, Arai AE, Lederman RJ, et al. 2007. Coronary artery aneurysms in patients with hyper IgE recurrent infection syndrome. *Clin. Immunol.* 122:255–58
105. Sharma A, Kumar S, Jagia P. 2018. Pulmonary artery pseudoaneurysm in hyper-IgE syndrome: rare complication with successful endovascular management. *Vasc. Endovasc. Surg.* 52:375–77
106. Takeuchi S, Wada K, Otani N, Nawashiro H. 2012. Multiple intracranial aneurysms associated with hyper-IgE syndrome. *Intern. Med.* 51:515–16
107. Freeman AF, Avila EM, Shaw PA, Davis J, Hsu AP, et al. 2011. Coronary artery abnormalities in Hyper-IgE syndrome. *J. Clin. Immunol.* 31:338–45
108. Beziat V, Li J, Lin JX, Ma CS, Li P, et al. 2018. A recessive form of hyper-IgE syndrome by disruption of ZNF341-dependent STAT3 transcription and activity. *Sci. Immunol.* 3:eaat4956
109. Frey-Jakobs S, Hartberger JM, Fliegauf M, Bossen C, Wehmeyer ML, et al. 2018. ZNF341 controls STAT3 expression and thereby immunocompetence. *Sci. Immunol.* 3:eaat4941
110. Schwerd T, Twigg SRF, Aschenbrenner D, Manrique S, Miller KA, et al. 2017. A biallelic mutation in *IL6ST* encoding the GP130 co-receptor causes immunodeficiency and craniosynostosis. *J. Exp. Med.* 214:2547–62
111. Shahin T, Aschenbrenner D, Cagdas D, Bal SK, Conde CD, et al. 2019. Selective loss of function variants in *IL6ST* cause Hyper-IgE syndrome with distinct impairments of T-cell phenotype and function. *Haematologica* 104:609–21
112. Ozaki K, Spolski R, Feng CG, Qi CF, Cheng J, et al. 2002. A critical role for IL-21 in regulating immunoglobulin production. *Science* 298:1630–34
113. Pesce J, Kaviratne M, Ramalingam TR, Thompson RW, Urban JF Jr., et al. 2006. The IL-21 receptor augments Th2 effector function and alternative macrophage activation. *J. Clin. Investig.* 116:2044–55

114. Neveu WA, Allard JB, Dienz O, Wargo MJ, Ciliberto G, et al. 2009. IL-6 is required for airway mucus production induced by inhaled fungal allergens. *J. Immunol.* 183:1732–38
115. Suto A, Nakajima H, Hirose K, Suzuki K, Kagami S, et al. 2002. Interleukin 21 prevents antigen-induced IgE production by inhibiting germ line C ϵ transcription of IL-4-stimulated B cells. *Blood* 100:4565–73
116. Kishida T, Hiromura Y, Shin-Ya M, Asada H, Kuriyama H, et al. 2007. IL-21 induces inhibitor of differentiation 2 and leads to complete abrogation of anaphylaxis in mice. *J. Immunol.* 179:8554–61
117. Avery DT, Ma CS, Bryant VL, Santner-Nanan B, Nanan R, et al. 2008. STAT3 is required for IL-21-induced secretion of IgE from human naive B cells. *Blood* 112:1784–93
118. Kotlarz D, Zietara N, Milner JD, Klein C. 2014. Human IL-21 and IL-21R deficiencies: two novel entities of primary immunodeficiency. *Curr. Opin. Pediatr.* 26:704–12
119. Kotlarz D, Zietara N, Uzel G, Weidemann T, Braun CJ, et al. 2013. Loss-of-function mutations in the IL-21 receptor gene cause a primary immunodeficiency syndrome. *J. Exp. Med.* 210:433–43
120. Palomares O, Martin-Fontecha M, Lauener R, Traidl-Hoffmann C, Cavkaytar O, et al. 2014. Regulatory T cells and immune regulation of allergic diseases: roles of IL-10 and TGF- β . *Genes Immun.* 15:511–20
121. Engelhardt KR, Shah N, Faizura-Yeop I, Kocacik Uygun DF, Frede N, et al. 2013. Clinical outcome in IL-10- and IL-10 receptor-deficient patients with or without hematopoietic stem cell transplantation. *J. Allergy Clin. Immunol.* 131:825–30
122. Spencer S, Kostel Bal S, Egner W, Lango Allen H, Raza SI, et al. 2019. Loss of the interleukin-6 receptor causes immunodeficiency, atopy, and abnormal inflammatory responses. *J. Exp. Med.* 216:1986–98
123. Ferreira RC, Freitag DF, Cutler AJ, Howson JM, Rainbow DB, et al. 2013. Functional *IL6R* 358Ala allele impairs classical IL-6 receptor signaling and influences risk of diverse inflammatory diseases. *PLOS Genet.* 9:e1003444
124. Wang Y, Hu H, Wu J, Zhao X, Zhen Y, et al. 2016. The *IL6R* gene polymorphisms are associated with sIL-6R, IgE and lung function in Chinese patients with asthma. *Gene* 585:51–57
125. Esparza-Gordillo J, Schaarschmidt H, Liang L, Cookson W, Bauerfeind A, et al. 2013. A functional IL-6 receptor (*IL6R*) variant is a risk factor for persistent atopic dermatitis. *J. Allergy Clin. Immunol.* 132:371–77
126. Puel A, Picard C, Lorrot M, Pons C, Chrabieh M, et al. 2008. Recurrent staphylococcal cellulitis and subcutaneous abscesses in a child with autoantibodies against IL-6. *J. Immunol.* 180:647–54
127. Ma CA, Xi L, Cauff B, DeZure A, Freeman AF, et al. 2017. Somatic STAT5b gain-of-function mutations in early onset nonclonal eosinophilia, urticaria, dermatitis, and diarrhea. *Blood* 129:650–53
128. Ando T, Xiao W, Gao P, Namiranian S, Matsumoto K, et al. 2014. Critical role for mast cell Stat5 activity in skin inflammation. *Cell Rep.* 6:366–76
129. Kawakami T, Ando T, Kawakami Y. 2015. Hypothetical atopic dermatitis-myeloproliferative neoplasm syndrome. *Front. Immunol.* 6:434
130. Bandapalli OR, Schuessele S, Kunz JB, Rausch T, Stutz AM, et al. 2014. The activating STAT5B N642H mutation is a common abnormality in pediatric T-cell acute lymphoblastic leukemia and confers a higher risk of relapse. *Haematologica* 99:e188–92
131. Kucuk C, Jiang B, Hu X, Zhang W, Chan JK, et al. 2015. Activating mutations of STAT5B and STAT3 in lymphomas derived from $\gamma\delta$ -T or NK cells. *Nat. Commun.* 6:6025
132. Rajala HL, Eldfors S, Kuusanmaki H, van Adrichem AJ, Olson T, et al. 2013. Discovery of somatic STAT5b mutations in large granular lymphocytic leukemia. *Blood* 121:4541–50
133. Cross NCP, Hoade Y, Tapper WJ, Carreno-Tarragona G, Fanelli T, et al. 2019. Recurrent activating STAT5B N642H mutation in myeloid neoplasms with eosinophilia. *Leukemia* 33:415–25
134. Del Bel KL, Ragotte RJ, Saferali A, Lee S, Vercauteren SM, et al. 2017. JAK1 gain-of-function causes an autosomal dominant immune dysregulatory and hypereosinophilic syndrome. *J. Allergy Clin. Immunol.* 139:2016–20.e5
135. Minegishi Y, Saito M, Morio T, Watanabe K, Agematsu K, et al. 2006. Human tyrosine kinase 2 deficiency reveals its requisite roles in multiple cytokine signals involved in innate and acquired immunity. *Immunity* 25:745–55
136. Kreins AY, Ciancanelli MJ, Okada S, Kong XF, Ramirez-Alejo N, et al. 2015. Human TYK2 deficiency: mycobacterial and viral infections without hyper-IgE syndrome. *J. Exp. Med.* 212:1641–62

137. Rawson R, Yang T, Newbury RO, Aquino M, Doshi A, et al. 2016. TGF- β 1-induced PAI-1 contributes to a profibrotic network in patients with eosinophilic esophagitis. *J. Allergy Clin. Immunol.* 138:791–800.e4
138. Balzar S, Chu HW, Silkoff P, Cundall M, Trudeau JB, et al. 2005. Increased TGF- β 2 in severe asthma with eosinophilia. *J. Allergy Clin. Immunol.* 115:110–17
139. Kotlarz D, Marquardt B, Baroy T, Lee WS, Konnikova L, et al. 2018. Human TGF- β 1 deficiency causes severe inflammatory bowel disease and encephalopathy. *Nat. Genet.* 50:344–48
140. Kinoshita A, Saito T, Tomita H, Makita Y, Yoshida K, et al. 2000. Domain-specific mutations in *TGFB1* result in Camurati-Engelmann disease. *Nat. Genet.* 26:19–20
141. Abonia JP, Wen T, Stucke EM, Grotjan T, Griffith MS, et al. 2013. High prevalence of eosinophilic esophagitis in patients with inherited connective tissue disorders. *J. Allergy Clin. Immunol.* 132:378–86
142. Morgan AW, Pearson SB, Davies S, Gooi HC, Bird HA. 2007. Asthma and airways collapse in two heritable disorders of connective tissue. *Ann. Rheum. Dis.* 66:1369–73
143. Loeys BL, Schwarze U, Holm T, Callewaert BL, Thomas GH, et al. 2006. Aneurysm syndromes caused by mutations in the TGF- β receptor. *N. Engl. J. Med.* 355:788–98
144. Dewan AK, Tomlinson RE, Mitchell S, Goh BC, Yung RM, et al. 2015. Dysregulated TGF- β signaling alters bone microstructure in a mouse model of Loeys-Dietz syndrome. *J. Orthop. Res.* 33:1447–54
145. Frischmeyer-Guerrero PA, Guerrero AL, Oswald G, Chichester K, Myers L, et al. 2013. TGF β receptor mutations impose a strong predisposition for human allergic disease. *Sci. Transl. Med.* 5:195ra94
146. Dietz HC, Cutting GR, Pyeritz RE, Maslen CL, Sakai LY, et al. 1991. Marfan syndrome caused by a recurrent de novo missense mutation in the fibrillin gene. *Nature* 352:337–39
147. Schalkwijk J, Zweers MC, Steijlen PM, Dean WB, Taylor G, et al. 2001. A recessive form of the Ehlers-Danlos syndrome caused by tenascin-X deficiency. *N. Engl. J. Med.* 345:1167–75
148. Lindsay ME, Dietz HC. 2014. The genetic basis of aortic aneurysm. *Cold Spring Harb. Perspect. Med.* 4:a015909
149. Lyons JJ, Liu Y, Ma CA, Yu X, O'Connell MP, et al. 2017. ERBIN deficiency links STAT3 and TGF- β pathway defects with atopy in humans. *J. Exp. Med.* 214:669–80. Correction. 2017. *J. Exp. Med.* 214:1201
150. Chen C, Akiyama K, Wang D, Xu X, Li B, et al. 2015. mTOR inhibition rescues osteopenia in mice with systemic sclerosis. *J. Exp. Med.* 212:73–91
151. Hershey GK, Friedrich MF, Esswein LA, Thomas ML, Chatila TA. 1997. The association of atopy with a gain-of-function mutation in the α subunit of the interleukin-4 receptor. *N. Engl. J. Med.* 337:1720–25
152. Oetjen LK, Mack MR, Feng J, Whelan TM, Niu H, et al. 2017. Sensory neurons co-opt classical immune signaling pathways to mediate chronic itch. *Cell* 171:217–28.e13
153. Brunkow ME, Jeffery EW, Hjerrild KA, Paepers B, Clark LB, et al. 2001. Disruption of a new forkhead/winged-helix protein, scurfin, results in the fatal lymphoproliferative disorder of the scurfy mouse. *Nat. Genet.* 27:68–73
154. Bennett CL, Christie J, Ramsdell F, Brunkow ME, Ferguson PJ, et al. 2001. The immune dysregulation, polyendocrinopathy, enteropathy, X-linked syndrome (IPEX) is caused by mutations of FOXP3. *Nat. Genet.* 27:20–21
155. Wildin RS, Ramsdell F, Peake J, Faravelli F, Casanova JL, et al. 2001. X-linked neonatal diabetes mellitus, enteropathy and endocrinopathy syndrome is the human equivalent of mouse scurfy. *Nat. Genet.* 27:18–20
156. Chatila TA, Blaeser F, Ho N, Lederman HM, Voulgaropoulos C, et al. 2000. *JM2*, encoding a fork head-related protein, is mutated in X-linked autoimmunity-allergic dysregulation syndrome. *J. Clin. Investig.* 106:R75–81
157. Ramsdell F, Ziegler SF. 2014. FOXP3 and scurfy: how it all began. *Nat. Rev. Immunol.* 14:343–49
158. Caudy AA, Reddy ST, Chatila T, Atkinson JP, Verbsky JW. 2007. CD25 deficiency causes an immune dysregulation, polyendocrinopathy, enteropathy, X-linked-like syndrome, and defective IL-10 expression from CD4 lymphocytes. *J. Allergy Clin. Immunol.* 119:482–87
159. Nadeau K, Hwa V, Rosenfeld RG. 2011. STAT5b deficiency: an unsuspected cause of growth failure, immunodeficiency, and severe pulmonary disease. *J. Pediatr.* 158:701–8
160. Kanai T, Jenks J, Nadeau KC. 2012. The STAT5b pathway defect and autoimmunity. *Front. Immunol.* 3:234

161. Josefowicz SZ, Niec RE, Kim HY, Treuting P, Chinen T, et al. 2012. Extrathymically generated regulatory T cells control mucosal T_H2 inflammation. *Nature* 482:395–99
162. Van Gool F, Nguyen MLT, Mumbach MR, Satpathy AT, Rosenthal WL, et al. 2019. A mutation in the transcription factor Foxp3 drives T helper 2 effector function in regulatory T cells. *Immunity* 50:362–77.e6
163. Sandilands A, Sutherland C, Irvine AD, McLean WH. 2009. Filaggrin in the frontline: role in skin barrier function and disease. *J. Cell Sci.* 122:1285–94
164. Smith FJ, Irvine AD, Terron-Kwiatkowski A, Sandilands A, Campbell LE, et al. 2006. Loss-of-function mutations in the gene encoding filaggrin cause ichthyosis vulgaris. *Nat. Genet.* 38:337–42
165. Sandilands A, Terron-Kwiatkowski A, Hull PR, O'Regan GM, Clayton TH, et al. 2007. Comprehensive analysis of the gene encoding filaggrin uncovers prevalent and rare mutations in ichthyosis vulgaris and atopic eczema. *Nat. Genet.* 39:650–54
166. Irvine AD, McLean WH, Leung DY. 2011. Filaggrin mutations associated with skin and allergic diseases. *N. Engl. J. Med.* 365:1315–27
167. Howell MD, Kim BE, Gao P, Grant AV, Boguniewicz M, et al. 2007. Cytokine modulation of atopic dermatitis filaggrin skin expression. *J. Allergy Clin. Immunol.* 120:150–55
168. Hvid M, Vestergaard C, Kemp K, Christensen GB, Deleuran B, Deleuran M. 2011. IL-25 in atopic dermatitis: a possible link between inflammation and skin barrier dysfunction? *J. Investig. Dermatol.* 131:150–57
169. Kottke MD, Delva E, Kowalczyk AP. 2006. The desmosome: cell science lessons from human diseases. *J. Cell Sci.* 119:797–806
170. Jonca N, Leclerc EA, Caubet C, Simon M, Guerrin M, Serre G. 2011. Corneodesmosomes and corneodesmosin: from the stratum corneum cohesion to the pathophysiology of genodermatoses. *Eur. J. Dermatol.* 21(Suppl. 2):35–42
171. Oji V, Eckl KM, Aufvenne K, Natebus M, Tarinski T, et al. 2010. Loss of corneodesmosin leads to severe skin barrier defect, pruritus, and atopy: unraveling the peeling skin disease. *Am. J. Hum. Genet.* 87:274–81
172. McAleer MA, Pohler E, Smith FJ, Wilson NJ, Cole C, et al. 2015. Severe dermatitis, multiple allergies, and metabolic wasting syndrome caused by a novel mutation in the N-terminal plaklin domain of desmoplakin. *J. Allergy Clin. Immunol.* 136:1268–76
173. Samuelov L, Sarig O, Harmon RM, Rapaport D, Ishida-Yamamoto A, et al. 2013. Desmoglein 1 deficiency results in severe dermatitis, multiple allergies and metabolic wasting. *Nat. Genet.* 45:1244–48
174. Ishida-Yamamoto A, Deraison C, Bonnart C, Bitoun E, Robinson R, et al. 2005. LEKTI is localized in lamellar granules, separated from KLK5 and KLK7, and is secreted in the extracellular spaces of the superficial stratum granulosum. *J. Investig. Dermatol.* 124:360–66
175. Chavanas S, Bodemer C, Rochat A, Hamel-Teillac D, Ali M, et al. 2000. Mutations in *SPINK5*, encoding a serine protease inhibitor, cause Netherton syndrome. *Nat. Genet.* 25:141–42
176. Jungersted JM, Scheer H, Mempel M, Baurecht H, Cifuentes L, et al. 2010. Stratum corneum lipids, skin barrier function and filaggrin mutations in patients with atopic eczema. *Allergy* 65:911–18
177. Kezic S, O'Regan GM, Lutter R, Jakasa I, Koster ES, et al. 2012. Filaggrin loss-of-function mutations are associated with enhanced expression of IL-1 cytokines in the stratum corneum of patients with atopic dermatitis and in a murine model of filaggrin deficiency. *J. Allergy Clin. Immunol.* 129:1031–39.e1
178. Miajlovic H, Fallon PG, Irvine AD, Foster TJ. 2010. Effect of filaggrin breakdown products on growth of and protein expression by *Staphylococcus aureus*. *J. Allergy Clin. Immunol.* 126:1184–90.e3
179. Simpson EL, Chalmers JR, Hanifin JM, Thomas KS, Cork MJ, et al. 2014. Emollient enhancement of the skin barrier from birth offers effective atopic dermatitis prevention. *J. Allergy Clin. Immunol.* 134:818–23
180. Kaplan AP, Beaven MA. 1976. In vivo studies of the pathogenesis of cold urticaria, cholinergic urticaria, and vibration-induced swelling. *J. Investig. Dermatol.* 67:327–32
181. Ombrello MJ, Remmers EF, Sun G, Freeman AF, Datta S, et al. 2012. Cold urticaria, immunodeficiency, and autoimmunity related to PLCG2 deletions. *N. Engl. J. Med.* 366:330–38
182. Gandhi C, Healy C, Wanderer AA, Hoffman HM. 2009. Familial atypical cold urticaria: description of a new hereditary disease. *J. Allergy Clin. Immunol.* 124:1245–50

183. Aderibigbe OM, Priel DL, Lee CC, Ombrello MJ, Prajapati VH, et al. 2015. Distinct cutaneous manifestations and cold-induced leukocyte activation associated with PLCG2 mutations. *JAMA Dermatol.* 151:627–34
184. Schade A, Walliser C, Wist M, Haas J, Vatter P, et al. 2016. Cool-temperature-mediated activation of phospholipase C- γ_2 in the human hereditary disease PLAID. *Cell Signal.* 28:1237–51
185. Wang J, Sohn H, Sun G, Milner JD, Pierce SK. 2014. The autoinhibitory C-terminal SH2 domain of phospholipase C- γ_2 stabilizes B cell receptor signalosome assembly. *Sci. Signal.* 7:ra89
186. Aksentijevich I, Putnam CD, Remmers EF, Mueller JL, Le J, et al. 2007. The clinical continuum of cryopyrinopathies: novel CIAS1 mutations in North American patients and a new cryopyrin model. *Arthritis Rheum.* 56:1273–85
187. Boyden SE, Desai A, Cruse G, Young ML, Bolan HC, et al. 2016. Vibratory urticaria associated with a missense variant in ADGRE2. *N. Engl. J. Med.* 374:656–63
188. Huang YS, Chiang NY, Hu CH, Hsiao CC, Cheng KF, et al. 2012. Activation of myeloid cell-specific adhesion class G protein-coupled receptor EMR2 via ligation-induced translocation and interaction of receptor subunits in lipid raft microdomains. *Mol. Cell. Biol.* 32:1408–20
189. Lyons JJ, Yu X, Hughes JD, Le QT, Jamil A, et al. 2016. Elevated basal serum tryptase identifies a multisystem disorder associated with increased TPSAB1 copy number. *Nat. Genet.* 48:1564–69
190. Lyons JJ, Sun G, Stone KD, Nelson C, Wisch L, et al. 2014. Mendelian inheritance of elevated serum tryptase associated with atopy and connective tissue abnormalities. *J. Allergy Clin. Immunol.* 133:1471–74
191. Ui H, Andoh T, Lee JB, Nojima H, Kuraishi Y. 2006. Potent pruritogenic action of tryptase mediated by PAR-2 receptor and its involvement in anti-pruritic effect of nafamostat mesilate in mice. *Eur. J. Pharmacol.* 530:172–78
192. Sommerhoff CP. 2001. Mast cell tryptases and airway remodeling. *Am. J. Respir. Crit. Care Med.* 164:S52–58
193. Doong JC, Chichester K, Oliver ET, Schwartz LB, Saini SS. 2017. Chronic idiopathic urticaria: systemic complaints and their relationship with disease and immune measures. *J. Allergy Clin. Immunol. Pract.* 5:1314–18
194. Fellingner C, Hemmer W, Wohrl S, Sesztak-Greinecker G, Jarisch R, Wantke F. 2014. Clinical characteristics and risk profile of patients with elevated baseline serum tryptase. *Allergol. Immunopathol.* 42:544–52
195. Kucharewicz I, Bodzenta-Lukaszyk A, Szymanski W, Mroczko B, Szmitkowski M. 2007. Basal serum tryptase level correlates with severity of hymenoptera sting and age. *J. Investig. Allergol. Clin. Immunol.* 17:65–69
196. Sahiner UM, Yavuz ST, Buyuktiryaki B, Cavkaytar O, Yilmaz EA, et al. 2014. Serum basal tryptase may be a good marker for predicting the risk of anaphylaxis in children with food allergy. *Allergy* 69:265–68
197. Valent P, Akin C, Metcalfe DD. 2017. Mastocytosis: 2016 updated WHO classification and novel emerging treatment concepts. *Blood* 129:1420–27
198. Le QT, Lyons JJ, Naranjo AN, Olivera A, Lazarus RA, et al. 2019. Impact of naturally forming human α/β -tryptase heterotetramers in the pathogenesis of hereditary α -tryptasemia. *J. Exp. Med.* 216:2348–61
199. Bernth-Jensen JM, Holm M, Christiansen M. 2016. Neonatal-onset T⁻B⁻NK⁺ severe combined immunodeficiency and neutropenia caused by mutated phosphoglucomutase 3. *J. Allergy Clin. Immunol.* 137:321–24
200. Sassi A, Lazaroski S, Wu G, Haslam SM, Fliegau M, et al. 2014. Hypomorphic homozygous mutations in phosphoglucomutase 3 (PGM3) impair immunity and increase serum IgE levels. *J. Allergy Clin. Immunol.* 133:1410–19.e13
201. Stray-Pedersen A, Backe PH, Sorte HS, Morkrid L, Chokshi NY, et al. 2014. PGM3 mutations cause a congenital disorder of glycosylation with severe immunodeficiency and skeletal dysplasia. *Am. J. Hum. Genet.* 95:96–107
202. Zhang Y, Yu X, Ichikawa M, Lyons JJ, Datta S, et al. 2014. Autosomal recessive phosphoglucomutase 3 (PGM3) mutations link glycosylation defects to atopy, immune deficiency, autoimmunity, and neurocognitive impairment. *J. Allergy Clin. Immunol.* 133:1400–9.e5

203. Ben-Khemis L, Mekki N, Ben-Mustapha I, Rouault K, Mellouli F, et al. 2017. A founder mutation underlies a severe form of phosphoglutamase 3 (PGM3) deficiency in Tunisian patients. *Mol. Immunol.* 90:57–63
204. Carlson RJ, Bond MR, Hutchins S, Brown Y, Wolfe LA, et al. 2017. Detection of phosphoglucomutase-3 deficiency by lectin-based flow cytometry. *J. Allergy Clin. Immunol.* 140:291–94.e4
205. Wu G, Hitchen PG, Panico M, North SJ, Barbouche MR, et al. 2016. Glycoproteomic studies of IgE from a novel hyper IgE syndrome linked to PGM3 mutation. *Glycoconj. J.* 33:447–56
206. Tanoue A, Endo F, Kitano A, Matsuda I. 1990. A single nucleotide change in the prolidase gene in fibroblasts from two patients with polypeptide positive prolidase deficiency: expression of the mutant enzyme in NIH 3T3 cells. *J. Clin. Investig.* 86:351–55
207. Fukumura A, Asaka T, Kasakura H, Doshita T, Chen W, et al. 2009. [Prolidase deficiency with various clinical conditions including hyper-IgE and multiple lung bulla formation]. *Nippon Naika Gakkai Zasshi* 98:150–52 (In Japanese)
208. Hershkovitz T, Hassoun G, Indelman M, Shlush LI, Bergman R, et al. 2006. A homozygous missense mutation in *PEPD* encoding peptidase D causes prolidase deficiency associated with hyper-IgE syndrome. *Clin. Exp. Dermatol.* 31:435–40
209. Lopes I, Marques L, Neves E, Silva A, Taveira M, et al. 2002. Prolidase deficiency with hyperimmunoglobulin E: a case report. *Pediatr. Allergy Immunol.* 13:140–42
210. Viglio S, Annovazzi L, Conti B, Genta I, Perugini P, et al. 2006. The role of emerging techniques in the investigation of prolidase deficiency: from diagnosis to the development of a possible therapeutical approach. *J. Chromatogr. B Analyt. Technol. Biomed. Life Sci.* 832:1–8
211. Lubick KJ, Robertson SJ, McNally KL, Freedman BA, Rasmussen AL, et al. 2015. Flavivirus antagonism of type I interferon signaling reveals prolidase as a regulator of IFNAR1 surface expression. *Cell Host Microbe* 18:61–74